# HATCHERY AND GENETIC MANAGEMENT PLAN (HGMP)

**Hatchery Program:** 

North Toutle Hatchery Type-S Coho (Integrated)

Species or Hatchery Stock:

Type-S Coho (*Oncorhynchus kisutch*)
Toutle River Stock

**Agency/Operator:** 

Washington Department of Fish and Wildlife

Watershed and Region:

Cowlitz River System/ Toutle Sub-Basin/ Lower Columbia River

**Date Submitted:** 

August 19, 2014

**Date Last Updated:** 

#### **Executive Summary**

The Washington Department of Fish and Wildlife is submitting a Hatchery and Genetic Management Plan (HGMP) for the North Toutle Hatchery Type-S Coho program to the National Marine Fisheries (NMFS) for consultation under Section 10(a)(1)(A) or 4(d) of the Endangered Species Act (ESA). NMFS will use the information in this HGMP to evaluate the hatchery impacts on salmon and steelhead listed under the ESA. The primary goal of an HGMP is to devise biologically-based hatchery management strategies that ensure the conservation and recovery of salmon and steelhead populations. This HGMP focuses on the implementation of hatchery reform actions adopted by the Washington Fish and Wildlife Commission Policy on Hatchery and Fishery Reform C-3619.

The purpose of the program is to produce Toutle River coho for sustainable escapement to the watershed, while providing recreational fisheries. Program fish will be produced at the North Toutle Hatchery, located on the Green River (WRIA 26.0323), tributary to the North Fork Toutle River (WRIA 26.0314) in the Cowlitz Basin. The program will annually release 150,000 yearlings to the Toutle River. In addition, this facility provides eggs for the Deep River Net Pen SAFE program (see Deep River Net Pen SAFE Coho HGMP).

This Type-S coho HGMP is built around the principles and recommendations of the Hatchery Scientific Review Group (HSRG). These principles and recommendations represent the best science available for operating hatchery facilities consistent with the conservation of salmonid species. The program has been operated as a "integrated type" program, as defined by the HSRG, since 2006. An "integrated" program is one in which natural-origin individuals are used in the hatchery broodstocks. Integration is achieved by using up to 30% of the returning adult natural-origin Type-S coho (distinguished by an intact adipose fin) returning to the North Toutle Hatchery trap on the Green River (RKm 0.8) from September through October. All fish released through this hatchery program have been 100% adipose fin-clipped (AD) since brood year 1995; release year 1997, of these, 20% of the on-station releases are also released AD+coded-wire tagged (CWT).

The Lower Columbia River coho are listed as "Threatened" under the ESA. The ESU includes the North Toutle River Hatchery Program.

#### **Broodstock Collection:**

The broodstock has been derived from 100% natural-origin stock returning to the North Fork Toutle/Green Rivers over the last three years. The current egg-take goal is 200,000 at North Toutle Hatchery; around 70 adult pairs may be collected. Surplus hatchery fish above broodstock needs may be passed above the hatchery, at a ratio designed to meet Primary population guidelines, as determined by Fish Management. Additional surplus may be donated to local food bank organizations or used for nutrient enhancement.

#### Harvest:

Total annual harvest is dependent on management response to annual abundance in *Pacific Salmon Commission* (PSC - U.S./Canada), *Pacific Fishery Management Council* (PFMC - U.S. ocean), and *Columbia River Compact* forums. WDFW has also received authorization for tributary, Columbia River mainstem, and ocean fisheries; the combined harvest rates in the *Fisheries Management and Evaluation Plan* (FMEP), *Columbia River Fish Management Plan* (CRFMP), and ocean fisheries are reviewed annually in the North of Falcon process. The *U.S. v Oregon* Technical Advisory Committee (TAC) has prepared Biological Assessments (BAs) for combined fisheries based on relevant *U.S. v Oregon* management plans and agreements. The current BA concerns Columbia River treaty Indian and non-Indian fisheries, as described in the "2008–2017 *U.S. v Oregon* Management Agreement for upriver Chinook, sockeye, steelhead, coho, and white sturgeon" (2008–2017 MA).

Hatchery coho can contribute significantly to the lower Columbia River gill net fishery; commercial harvest of early-returning (Type-S) coho is constrained by fall Chinook and Sandy River coho

management; commercial harvest of late-returning (Type-N) coho is focused in October during the peak abundance of hatchery-origin Type-N coho. A substantial estuary sport fishery exists between Buoy 10 and the Astoria-Megler Bridge; majority of the catch is hatchery-origin Type-S coho, but the hatchery-origin Type-S coho harvest can also be substantial.

Due to limitations that not all fish can be accounted for as being harvested or as back-to-rack counts, smolt-to-adult survival rates (SAR) are likely underestimated. Based on the average SAR of 2.39% for brood years 2001-2005, and a programmed release goal of 150,000 yearlings, the estimated production goal would be 3,585 adults.

#### **Monitoring and Evaluation:**

Performance indicators for harvest will be accomplished by continuing mass-marking (adipose fin-clip); CWT recoveries help determine stray rate contributions on spawning grounds by watersheds close in proximity to this program's release vicinity.

#### **Operation and Maintenance of Hatchery Facilities:**

North Toutle Hatchery has water rights to divert water at a rate of 48 cfs from the Green River The intake structures are in compliance with state and federal guidelines, but do not meet the current criteria. WDFW has requested funding for scoping, design, and construction work of a new intake system. Upstream passage is blocked by the temporary barrier weir during the broodstock collection period. The return water systems operate under a National Pollutant Discharge Elimination System (NPDES) permit.

#### 1 SECTION 1. GENERAL PROGRAM DESCRIPTION

#### 1.1 Name of hatchery or program.

North Toutle Hatchery Type-S Coho

#### 1.2 Species and population (or stock) under propagation, and ESA status.

Toutle River Type-S Coho (*Oncorhynchus kisutch*)

ESA Status: "Threatened" June 28, 2005 (70FR37160); reaffirmed on August 15, 2011 (76 FR 50448).

#### 1.3 Responsible organization and individuals

Hatchery Operations Staff Lead Contact

Name (and title): Mark Johnson, Hatcheries Operations and Complex Manager

**Agency or Tribe:** Washington Department of Fish & Wildlife

**Address:** 165 Osprey Lane, Toledo WA 98591

**Telephone:** (360) 864-6135 **Fax:** (360) 864-6122

Email: Mark.Johnson@dfw.wa.gov

Fish Management Staff Lead Contact

Name (and title): Eric Kinne, Region 5 Hatchery Reform Coordinator

**Agency or Tribe:** Washington Dept. of Fish and Wildlife

Address: 2108 Grand Boulevard, Mail Stop: S-19, Vancouver, WA 98661-4624

**Telephone:** (360) 906-6747 **Fax:** (360) 906-6776

Email: Eric.Kinne@dfw.wa.gov

Other agencies, Tribes, co-operators, or organizations involved, including contractors, and extent of involvement in the program:

NOAA-National Marine Fisheries Service (NMFS) – Manager of Mitchell Act Funds

#### 1.4 Funding source, staffing level, and annual hatchery program operational costs.

**Funding Sources** Operation Information

Mitchell Act Full time equivalent staff – 2.76

Annual operating cost (dollars) - \$323,922

The above information for full-time equivalent staff and annual operating cost applies cumulatively to anadromous program facilities and cannot be broken out specifically by program.

#### 1.5 Location(s) of hatchery and associated facilities.

**Broodstock Source:** Toutle River Type-S (early-returning) coho

**Table 1.5.1:** Location of culturing phases, by facility.

Facility	Culturing Phase	Location
North Toutle	Broodstock collection,	Green River (WRIA 26.0323) at RM 0.81; tributary to North
Hatchery	Adult holding/spawning, Incubation, Rearing,	Fork Toutle River (WRIA 26.0314) at R.M. 11.5; tributary to the Toutle River (WRIA 26.0227) at R.M. 17.2; tributary
	Acclimation Acclimation	to the Cowlitz River (WRIA 26.0002) at R.M. 20.0;
		tributary to the Columbia River at R.M. 68.0, Lower
		Columbia River, Washington.

#### 1.6 Type of program.

Integrated Harvest/Conservation

#### 1.7 Purpose (Goal) of program.

Mitigation/Augmentation. The goal of this program is to provide maximum sport harvest under the mark-selective fishery regulations (retention of adipose-clipped fish only) while minimizing impacts to wild listed salmonids and steelhead. This program is a conservation/harvest program.

The proposed integrated strategy for this program is based on WDFW's assessment of the genetic characteristics of the hatchery and local natural population, the current and anticipated productivity of the habitat used by the populations, the potential for successfully implementing an isolated program, and NMFS' listing determination (August 15, 2011 76 FR 50448).

In addition, this facility provides HxH eggs (segregated program) for the BPA-funded Selected Area Fishery Enhancement (SAFE) program at Deep River Net Pens (see Deep River Net Pen SAFE Coho HGMP).

#### 1.8 Justification for the program.

The program is funded through the Mitchell Act via NOAA-NMFS for the purpose of mitigation for lost fish production due to development within the Columbia River Basin. WDFW protects listed fish and provides harvest opportunity on hatchery fish through the Lower Columbia River-Fish Management and Evaluation Plan (FMEP) (WDFW 2001). All mainstem and tributary salmon fisheries are managed as mark-selective (no wild retention) fisheries to minimize the impact on listed wild fish. The Mitchell Act programs are intended to support Northwest fishing economies – particularly coastal and Native American — that have relied on Columbia River production both before and after dam construction. Catches of hatchery fish sustain the economies of local communities while keeping incidental mortalities of ESA-Listed fish at approved levels. Value of hatchery production and benefit to local economies will be further increased by implementing fisheries that increase harvest of hatchery produced fish, as expected through implementation of the Lower Columbia Salmon Recovery Plan (LCSRP).

To minimize impact on listed fish by the North Toutle Hatchery Type-S Coho program and operations, the following risk aversions are included in this HGMP (**Table 1.8.1**).

**Table 1.8.1:** Summary of risk aversion measures for the North Toutle Type-N Coho program.

Potential Hazard	<b>HGMP Reference</b>	Risk Aversion Measures		
Water Withdrawal	4.1	Water rights are formalized through trust water right from the Department of Ecology. Monitoring and measurement of water usage is reported in monthly NPDES reports.		
Intake Screening	4.1	The intake screens are in compliance with state and federal guidelines (NMFS 1995, 1996), but do not meet the current <i>Anadromous Salmonid Passage Facility Design Criteria</i> (NMFS 2011). WDFW has secured funding in 2012 for scoping, design, and construction work of a new river intake system to meet NOAA-NMFS compliance (Mitchell Act Intake and Fish Passage Study Report 2003).		
Effluent Discharge	4.1	This facility operates under the "Upland Fin-Fish Hatching and Rearing" National Pollution Discharge Elimination System (NPDES) administered by the Washington Department of Ecology (DOE) - WAG 13-1010.		
Broodstock Collection & Adult Passage	7.9	All fish are mass marked prior to release. Broodstock collection and sorting procedures can quickly identify listed non-target listed fish, and if encountered, released per protocol to minimize impact as determined by WDFW		

		Region 5 staff.
		The intake facilities need repairs to provide compliant passage.
Disease Transmission	7.9, 10.11	The Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State (WDFW and WWTIT 1998, updated 2006) and the Fish Health Policy in the Columbia Basin details hatchery practices and operations designed to stop the introduction and/or spread of any diseases within the Columbia Basin. Also, Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries (Fish Health Policy Chapter 5, IHOT 1995).
Competition & Predation	2.2.3, 10.11	Fish are released at a time, size and the system and life history stage to foster rapid migration to marine waters, and to allow juvenile listed fish to grow to a size that reduces potential for predation.  Current risk aversions and future considerations are being reviewed and evaluated for further minimizing impacts to listed fish.

#### 1.9 List of program "Performance Standards".

See HGMP section 1.10. Standards and indicators are referenced from Northwest Power Planning Council (NPPC) Artificial Production Review (APR) (NPPC 2001).

#### 1.10 List of program "Performance Indicators", designated by "benefits" and "risks."

#### 1.10.1 "Performance Indicators" addressing benefits.

**Table 1.10.1.1:** "Performance Indicators" addressing benefits.

Benefits				
Performance Standard	Performance Indicator	Monitoring & Evaluation		
3.1.2 Program contributes to mitigation requirements. Program provides mitigation for lost fish production due to development within the Columbia River Basin.	Number of fish released by program returning, or caught, as applicable to given mitigation requirements.	Annually estimate survival and contribution for each brood year released.  This program provides mitigation for lost fish production due to development within the Columbia River Basin and contributes to a meaningful harvest in sport and commercial fisheries.		
3.1.3 Program addresses ESA responsibilities.	Program complies with Federal ESA-listed fish take authorizations for harvest and hatchery actions.	Compliance with ESA is managed with sport fishery regulations that minimize impacts to ESA-listed fish and are monitored by WDFW law enforcement officers. The FMEP outlines anticipated encounter rates and expected mortality rates for these fisheries. Creel surveys are being implemented to verify. Natural populations are monitored annually to assess trends and compare with goals. HGMP updated and re-submitted		

		to NOAA with significant changes or under permit agreement.
3.2.1 Fish produced for harvest are propagated and released in a manner enabling effective harvest, as described in all	Annual number of fish produced by program caught in all fisheries, including estimates of fish released.	Annually mass-mark hatchery releases to differentiate hatchery from natural-origin fish and record estimates of mark rate.
applicable fisheries management plans, while adequately minimizing by-catch of non-target species.		The external mark enables mark- selective fisheries, which can reduce directed harvest mortality on natural-origin fish.
		Harvest is regulated to meet appropriate biological assessment criteria. Agencies monitor harvests to provide up-to-date information.
		Estimate survival and contribution to fisheries for each brood year released.
3.3.1. Artificial propagation program contributes to an increasing number of spawners returning to natural spawning areas.	Annual number of naturally- produced adults or redds on the spawning grounds or selected natural production index areas.	Annually monitor and report returns to the hatchery and spawning grounds
3.3.2 Releases are sufficiently marked to allow statistically significant evaluation of program contribution to natural	Percentage of total hatchery releases are identifiable as hatchery-origin fish. Mass-mark (fin-clip, CWT, otolith-mark,	Annually monitor and report size, number, mass-mark quality (mark rate/tag rate) and date of all hatchery releases.
production, and to evaluate effects of the program on the local natural population.	other, etc., depending on species) production fish to identify them from naturally produced fish.	Annually sample returning fish for the mass-mark and CWT in fisheries and at the hatchery; monitor and report numbers of estimated hatchery (marked) and natural (unmarked) fish.
		Report CWT analysis to RMIS database.
3.4.1 Fish collected for broodstock are taken throughout the return or spawning period in proportions approximating the timing and age distribution of population from which	Temporal distribution of broodstock collection at point of collection.	Collect broodstock representatively and systematically throughout the early portion of the return (September through mid- November).
broodstock is taken.		Collect annual run timing, age and sex composition and spawning escapement timing data.
		Adhere to WDFW spawning guidelines (Seidel 1983; HSRG 2009).
3.5.5 Juveniles are released at fully-smolted stage to benefit juvenile to adult survival rates, and reduce the likelihood for	Level of smoltification (size, appearance, behavior, etc.) at release compared to WDFW rearing and release guidelines.	Monitor fish condition in the facilities throughout all rearing stages.  Annually monitor and record

residualism and negative ecological interactions with natural-origin fish.	Release type (forced, volitional, or direct).	size, number, and date of release.
3.6.1 The hatchery program uses standard scientific procedures to evaluate various aspects of artificial propagation.	Apply basic monitoring standards in the hatchery: food conversion rates, growth trajectories, mark/tag rate error, weight distribution (CV).	Collect annual run timing, age and sex composition data upon adult return.  Annually record growth rates, mark rate and size at release and release dates.  See also HGMP section 11 for program monitoring and evaluation.
3.8.3 Non-monetary societal benefits for which the program is designed are achieved.	Program is designed to help achieve the end goal of conserving and stabilizing natural salmon populations.	Long-term monitoring of system population will indicate success of program.

#### 1.10.2 "Performance Indicators" addressing risks.

**Table 1.10.2.1**: "Performance indicators" addressing risks.

Risks				
Performance Standard Performance Indicator Monitoring & Evaluation				
3.1.3 Program addresses ESA responsibilities	Program complies with Federal ESA-listed fish take authorizations for harvest and hatchery actions.	HGMP is updated to reflect any major changes in program and resubmitted to NOAA fisheries.		
		Program risks have been addressed in this HGMP through best available science hatchery management actions.		
		WDFW staff annually reviews Future Brood Document (FBD) for stock, size, number, date and location of releases from all production programs.		
		Monitor and record juvenile hatchery fish size, number, date of release and mass-mark quality; monitor contribution of hatchery adult fish to fisheries and escapement.		
3.2.1 Fish produced for harvest are produced and released in a manner enabling effective harvest, as described in all	Annual number of naturally- produced adults and/or redds on the spawning grounds or in selected natural production index	Production fish are mass-marked (adipose fin-clip) to allow for their differentiation from naturally-produced fish		
applicable fisheries management plans, while adequately minimizing by-catch of non-target species.	areas.  Number of marks released and estimated proportion of marks in out-migrant juveniles and returning adults on the spawning ground.	Monitor and record juvenile hatchery fish size, number, date of release and mass-mark (fin clips, tags, etc.) quality; monitor contribution of hatchery adult fish to fisheries and escapement.		
		Harvest is regulated to meet appropriate biological assessment criteria.		

		Agancies monitor horyests and
		Agencies monitor harvests and hatchery escapements to provide up-to-date information.
3.2.2 Release groups are sufficiently marked in a manner consistent with information needs and protocols to enable	Percentage of total hatchery releases are identifiable as hatchery-origin fish. Mass-mark (adipose-fin clip, CWT, otolith-	Annually monitor and report size, number, date of release and mass-mark quality (adipose finclip rate) of all hatchery releases.
determination of impacts to natural- and hatchery-origin fish in fisheries.	mark, etc., depending on species) produced fish to allow for their differentiation from naturally produced fish for selective fisheries.	Annually assess harvest of mass- marked hatchery fish based on CRC estimates and creel surveys.
3.3.2 Releases are sufficiently marked to allow statistically significant evaluation of program contribution to natural production and to evaluate effects of the	All hatchery production is identifiable in some manner (fin-marks, tags, otolith, etc.) consistent with information needs.	Annually monitor and record size, number, date of release and mass-mark quality (tag rate) of hatchery releases.
program on the local natural population.	needs.	Examine returning fish encountered for the mass-mark (CWT) at the hatchery and on the spawning ground. Annually record numbers of estimated hatchery (marked) and natural (unmarked).
3.4.1 Fish collected for broodstock are taken throughout the return or spawning period in proportions approximating the timing and age distribution of population from which broodstock is taken.	Temporal and age distribution of broodstock collected, compared to that of naturally-produced population at collection point.	Collect annual run timing, age and sex composition and return timing data.
3.4.3 Life history characteristics of the natural population do not change as a result of the hatchery	Life history characteristics are measured in adult and juvenile hatchery fish: return timing, age	Collect annual run timing, origin, and age and sex composition data.
program.	and sex composition, spawning timing, and size in returning hatchery adults; size, growth rates, and survival to release in juvenile production.	Annually monitor and record juvenile hatchery fish size, growth rates, number released, mass-mark/tag data, survival-to-release rates, and date of release.
	Life history patterns of juvenile and adult NOR are stable.	Examine returning fish for the mass-mark (fin-clips, CWTs) at broodstock collection points and on the spawning grounds.  Annually record and report numbers of estimated hatchery (marked) and natural (unmarked).
3.5.1 Patterns of genetic variation within and among natural populations do not change significantly as a result of artificial production.	Within and between populations, genetic structure is not affected by artificial production.	See HGMP section 11 for M&E information.
3.5.2 Collection of broodstock does not adversely impact the genetic diversity of the naturally-spawning population.	Total number of natural-origin spawners (if any) reaching the collection facility.	All on-station hatchery releases are identifiable in some manner (fin-marks, tags, etc.).

25 211.41	Timing of collection compared to overall run timing.	Collect annual run timing, origin, and age and sex composition data.  CWT data reported to RMIS.  Examine returning fish for the mass-mark (fin-clips, CWTs) at broodstock collection points and on the spawning grounds.  Annually record and report numbers of estimated hatchery (marked) and natural (unmarked).
3.5.3 Hatchery-origin adults in natural production areas do not negatively affect the total natural spawning population.	The ratio of observed and/or estimated total numbers of artificially-produced fish on natural spawning grounds, to total number of naturally-produced fish (pHOS).	Examine returning fish for the mass-mark (fin-clips, CWTs) at broodstock collection points and on the spawning grounds.  Annually record and report numbers of estimated hatchery (marked) and natural (unmarked).
3.5.4 Juveniles are released onstation, or after sufficient acclimation to maximize homing ability to intended return locations.	Location of release (on-station, acclimation pond, direct plant).  Release type (forced, volitional or direct stream release).  Proportion of adult returns to program's intended return location, compared to fisheries and artificial or natural production areas.	Examine returning fish for the mass-mark (fin-clips, CWTs) at broodstock collection points and on the spawning grounds.  Annually record and report numbers of estimated hatchery (marked) and natural (unmarked).  Annually record and report release information, including location, method and age class in hatchery data systems (WDFW Hatcheries Headquarters Database).
3.5.5 Juveniles are released at fully-smolted stage.  3.7.1 Hatchery facilities are operated in compliance with all applicable fish health guidelines and facility operation standards and protocols (IHOT, PNFHPC, Salmonid Disease Control Policy of the Fisheries Co-Managers of	Level of smoltification at release. Release type (forced, volitional or direct).  Annual reports indicating levels of compliance with applicable standards and criteria.  Periodic audits indicating level of compliance with applicable standards and criteria.	Annually monitor and record size, number, date of release and release type.  Pathologists from WDFW's Fish Health Section monitor program monthly. Exams performed at each life stage may include tests for virus, bacteria, parasites and/or pathological changes, as needed.
Washington State).		The program is operated consistent with the Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State (WDFW and WWTIT 1998, updated 2006), Fish Health Policy in the Columbia Basin, and Policies and Procedures for Columbia Basin Anadromous Salmonid

		Hatcheries (Fish Health Policy Chapter 5, IHOT 1995).
3.7.2 Effluent from hatchery facility will not detrimentally affect natural populations.	Discharge water quality compared to applicable water quality standards by NPDES permit.  WDFW water right permits	Flow and discharge reported in monthly NPDES report (see HGMP section 4.2).
	compliance.	
3.7.3 Water withdrawals and instream water diversion structures for artificial production facility operation will not prevent access to natural spawning areas, affect spawning behavior of natural populations, or impact juvenile rearing environment.	Water withdrawals compared to NMFS, USFWS and WDFW applicable passage and screening criteria for juveniles and adults.	Barrier and intake structure compliance assessed and needed fixes are prioritized (see HGMP section 4.2).
3.7.4 Releases do not introduce pathogens not already existing in the local populations, and do not significantly increase the levels of existing pathogens. Follow the Salmonid Disease Control Policy	Necropsies of fish to assess health, nutritional status, and culture conditions.	DFW Fish Health Section inspect adult broodstock yearly for pathogens and monitor juvenile fish on a monthly basis to assess health and detect potential disease problems.
of the Fisheries Co-Managers of Washington State (WDFW and WWTIT 1998, revised 2006).		A fish health database will be maintained to identify trends in fish health and disease and implement fish health management plans based on findings.
	Release and/or transfer exams for pathogens and parasites.	Examine fish 1 to 6 weeks prior to transfer or release, in accordance with the <i>Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State</i> (WDFW and WWTIT 1998, updated 2006).
	Inspection of adult broodstock for pathogens and parasites.	At spawning, lots of 60 adult broodstock are examined for pathogens.
	Inspection of off-station fish/eggs prior to transfer to hatchery for pathogens and parasites.	Controls of specific fish pathogens through eggs/fish movements are conducted in accordance to the Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State (WDFW and WWTIT 1998, updated 2006).
3.7.6 Adult broodstock collection operation does not significantly alter spatial and temporal distribution of any naturally-produced population.	Spatial and temporal spawning distribution of natural populations above and below broodstock collection site is currently compared to historic distribution.	Trap is checked daily. Non-target listed fish, when encountered, are returned to the river.
3.7.7 Weir/trapping operations do not result in significant stress, injury or mortality in natural	Mortality rates in trap.  Pre-spawning mortality rates of captured fish in the hatchery	Traps checked daily. Annually record and report abundances and observations of natural- origin

populations.	and/or after release.	fish at hatchery facilities.
3.7.8 Predation by artificially produced fish on naturally – produced fish does not significantly reduce numbers of natural fish.	Hatchery juveniles are raised to smolt-size and released from the hatchery at a time that fosters rapid migration downstream.	Hatchery smolt release size and time are monitored to quantify/minimize predation effects on naturally-origin salmon and steelhead (Sharpe et al. 2008).
3.8.1 Cost of program operation does not exceed the net economic value of fisheries in dollars per fish for all fisheries targeting this population.	Total cost of operation.	Compare annual operational cost of program to calculated fishery contribution value (Wegge 2009).
3.8.2. Juvenile production costs are comparable to or less than other regional programs designed for similar objectives.	Total cost of program operation.	Annually monitor and report feed costs and fish health actions.

#### 1.11 Expected size of program.

## 1.11.1 Proposed annual broodstock collection level (maximum number of adult fish).

*Integrated program.* Around 70 adult pairs, not including jacks, are needed to achieve the established egg-take goal of 200,000 (FBD 2014) for the on-station program. This is based on an average fecundity of around 3,000 eggs/female, and a pre-spawning mortality of 10%.

*Segregated program.* A total of around 335 hatchery-origin adult pairs are needed to achieve the egg-take goal of 910,000 collected for the Selected Area Fisheries Enhancement (SAFE) releases from Deep River Net Pens (see Deep River Net Pens SAFE Coho HGMP).

**Table 1.11.1.1:** Egg-take goals for North Toutle Hatchery integrated and segregated program broodstocks.

Program	Egg-Take	Program Type	Location	Major Watershed
North Toutle on-station	200,000	Integrated	Green River	Cowlitz
Deep R net pen (SAFE program)	910,000	Segregated	Deep River	Grays-Elochoman

Source: Future Brood Document 2014.

## 1.11.2 Proposed annual fish release levels (maximum number) by life stage and location.

**Table 1.11.2.1:** Proposed annual fish release levels (maximum number) by life stage and location. North Toutle Hatchery on-station program.

Age Class	Max. No.	Release Date	Location	Major Watershed
Yearlings	150,000	May	Green River	Cowlitz

Source: Future Brood Document 2014.

See also Deep River Net Pens SAFE Coho HGMP.

### 1.12 Current program performance, including estimated smolt-to-adult survival rates, adult production levels, and escapement levels. Indicate the source of these data.

**Table 1.12.1:** Total releases and hatchery escapement of N. Toutle Type S Coho, 2002-2013.

Return	Total	Hatchery
Year	Release	Escapement
2002	986,490	22,769
2003	808,100	32,243

Average	546,615	15,937
2013	163,289	3,060
2012	155,310	2,498
2011	164,985	3,319
2010	147,232	10,344
2009	489,641	23,887
2008	829,666	33,797
2007	755,112	18,149
2006	691,881	7,493
2005	597,750	7,992
2004	769,926	25,693

Source: WDFW Hatcheries Headquarters Database 2014.

See also **Table 3.3.1.1**.

#### 1.13 Date program started (years in operation), or is expected to start.

The North Toutle Hatchery began operation in 1951. The Toutle River was planted with coho until the eruption of Mt. Saint Helens in 1980. Coho plants resumed post-eruption with brood year 1984, and rearing ponds were developed near the site of the original hatchery in 1985. The hatchery was renovated and resumed operations in 1991.

#### 1.14 Expected duration of program.

Program is on-going, with no plans for termination.

#### 1.15 Watersheds targeted by program.

Cowlitz Sub-Basin, Lower Columbia River.

### 1.16 Indicate alternative actions considered for attaining program goals, and reasons why those actions are not being proposed.

#### 1.16.1 Brief Overview of Key Issues.

Fish passage at North Toutle Hatchery is managed by the in-stream fish barrier and fish ladder. The WDFW Columbia River Fisheries Development (CRFD) 2003 *Intake and Fish Passage Study Report* indicates the passage at North Toutle is not in compliance with current passage criteria. This trap facility has several issues related to unsafe handling of adult listed fish. A complete investigation and comprehensive re-design is needed to accommodate a facility that can be installed and removed without putting machinery in the stream, as well as a trap facility that will sort, return to the stream, and or load fish with a water to water transfer method to cause no harm to hatchery or wild stocks. Adult sorting and handling in general is very hard on adult fish and routinely causes mortality that can be prevented with a modern sorting and handling system designed to cause the least harm possible to all fish handled.

#### 1.16.2 Potential Alternatives to the Current Program

Alternative 1: Eliminate the program: This action would reduce potential interaction with natural populations and eliminate potential impacts on other ESA-listed species. Currently this program supports popular sport fisheries in the lower Columbia River, and is consistent with the mitigation requirements.

Alternative 2: Use adults and acclimation ponds sited near appropriate habitat as release points. This would spread the returning adults out in the system and encourage them to utilize under escaped habitat.

#### 1.16.3 Potential Reforms and Investments

Reform/Investment 1: Modernize the Toutle Trap. A semi-automated sorting system would be comprised of the following: Construct a three pond system that allows for fish to enter a presorting pond volitionally. Fish would be sorted from this pond and either returned to stream above the weir, held for broodstock or surplussed. All ponds would be equipped with a crowder and a brail type system.

Reform/Investment 2: Fish passage at North Toutle Hatchery. The solution to the passage non-compliance is not one that can be solved by operating the fishway or hatchery water intake in a different manner. The solution lies in the construction of significant modification of the existing fishway/barrier or a completely new fishway/weir.

Reform/Investment 3: A non-compliant status also exists with the gravity intake for the rearing ponds. Screen slot width, approach velocities, sweep velocities, and floodwater invasion are the non-compliant issues. The solution to these issues will be a significant construction and possibly a complete rebuild.

Reform/Investment 4: Water supply system has pipeline problems and needs to be replaced form the gravity intake to the hatchery rearing ponds, as well as new riser supply piping.

#### 2 <u>SECTION 2. PROGRAM EFFECTS ON NMFS ESA-LISTED</u> <u>SALMONID POPULATIONS. (USFWS ESA-Listed Salmonid Species</u> and Non-Salmonid Species are addressed in Addendum A)

2.1 List all ESA permits or authorizations in hand for the hatchery program.

None currently. This HGMP is submitted to the NOAA Fisheries for ESA consultation and take prohibition exemption under ESA section 4(d) or 10.

- 2.2 Provide descriptions, status, and projected take actions and levels for NMFS ESA-listed natural populations in the target area.
  - 2.2.1 <u>Description of NMFS ESA-listed salmonid population(s) affected by the program.</u>
  - Identify the NMFS ESA-listed population(s) that will be <u>directly</u> affected by the program.

**Lower Columbia River coho** (*Oncorhynchus kisutch*). Identified as a candidate species on June 25, 1995 (60FR38011). Listed as threatened on June 28, 2005 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

- Identify the NMFS ESA-listed population(s) that may be incidentally affected by the program.

**Lower Columbia River steelhead** (*Oncorhynchus mykiss*). Listed as a threatened species on March 19, 1998 (63FR13347); threatened status reaffirmed on January 5, 2006 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

**Lower Columbia River Chinook** (*Oncorhynchus tshawytscha*). Listed as "threatened" on March 24, 1999 (64FR14308); threatened status reaffirmed on June 28, 2005 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

**Columbia River chum salmon** (*Oncorhynchus keta*). Listed as threatened on March 25, 1999 (64FR14507); threatened status reaffirmed on June 28, 2005 (70FR37160); reaffirmed threatened by five-year status review, completed August 15, 2011 (76 FR 50448).

#### 2.2.2 Status of NMFS ESA-listed salmonid population(s) affected by the program.

- Describe the status of the listed natural population(s) relative to "critical" and "viable" population thresholds.

**Lower Columbia River Chinook**: In Washington, the LCR Chinook ESU includes all naturally spawned Chinook populations from the mouth of the Columbia to a transitional point between Washington and Oregon east of the Hood River and the White Salmon River, as well as fifteen artificial propagation programs. Excluded are Upper Columbia River bright hatchery stocks that spawn in the mainstem Columbia River below Bonneville Dam and in other tributaries upstream from the Sandy River to the Hood and White Salmon rivers (NMFS 2014 79FR20802).

Status: Today only two of 32 historical populations – the North Fork Lewis and Sandy late-fall populations – are considered viable. Most populations (26 out of 32) have a very low probability of persistence over the next 100 years, and some populations are extirpated, or nearly so. Five of the six strata fall significantly short of the Willamette- Lower Columbia Technical Recovery Team (WLC TRT) criteria for viability. One stratum – Cascade late fall – meets the WLC TRT criteria (Dornbush and Sihler 2013). Dam construction eliminated habitat for a number of populations leading to the extirpation of spring Chinook salmon populations in the Upper Cowlitz, Cispus, Tilton, North Fork Lewis, Big White Salmon rivers, and fall Chinook populations in the Upper Cowlitz and Big White Salmon rivers (SHIEER, NMFS 2004). Projects to allow access have been initiated in the Cowlitz and Lewis systems but these are not close to producing self-sustaining populations; Condit Dam on the Big White Salmon River was breached October 26, 2011. Based on the 2010 recovery plan analyses, all of the 14 Tule populations (Table 2.2.2.1) are considered very high risk except one that is considered at high risk. The modeling conducted in association with Tule harvest management suggests that three of the populations (Coweeman, Lewis and Washougal) are at a somewhat lower risk (LCFRB 2010).

**Table 2.2.2.1:** Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River Chinook populations.

		Ba	seline	viabil	ity		Prod.		Abundance	
Population	Contribution	A&P	s	D	Net	Obj.	target	Historical	Baseline	Target
Coast Fall										
Grays/Chinook	Contributing <sup>2</sup>	VL	Н	VL	$VL^2$	M+	+500%	800	<50	1,000
Eloch/Skam <sup>c</sup>	Primary	VL	Н	L	VL <sup>2</sup>	Н	+150%	3,000	<50	1,500
Mill/Aber/Germ	Primary <sup>1</sup>	VL	Н	L	$VL^2$	Н	+155%	2,500	50	900
Youngs Bay (OR)	Stabilizing	3	3	3	L	L	3	3	_3	3
Big Creek (OR) c	Contributing <sup>1</sup>	3	3	3	VL	L	3	3	3	3
Clatskanie (OR)	Primary	3	3	3	VL	Н	3	3	3	3
Scappoose (OR)	Primary <sup>1</sup>	3	3	3	L	н	3	3	3	3
Cascade Fall										
Lower Cowlitz <sup>c</sup>	Contributing	VL	Н	M	VL <sup>2</sup>	M+	+50%	24,000	500	3,000
Upper Cowlitz	Stabilizing	VL	VL	М	VL	VL		28,000	0	
Toutle <sup>c</sup>	Primary <sup>1</sup>	VL	Н	M	VL <sup>2</sup>	H+	+265%	11,000	<50	4,000
Coweeman <sup>G</sup>	Primary	VL	Н	Н	$VL^2$	H+	+80%	3,500	100	900
Kalama	Contributing <sup>2</sup>	VL	Н	М	$VL^2$	M	+110%	2,700	<50	500
Lewis <sup>G</sup>	Primary	VL	Н	Н	$VL^2$	H+	+280%	2,600	<50	1,500
Salmon	Stabilizing	VL	Н	М	VL	VL		n/a	<50	
Washougal	Primary	VL	Н	М	$VL^2$	H+	+190%	2,600	<50	1,200
Clackamas (OR) <sup>C</sup>	Contributing	3	3	3	VL	M	3	3	3	3
Sandy (OR)	Contributing 1	3	3	3	VL	M	3	3	3	3
Cascade L Fall										
Lewis NF <sup>C,G</sup>	Primary	VH	Н	Н	VH1	VH	0%	23,000	7,300	7,300
Sandy (OR) <sup>c,g</sup>	Primary	3	3	3	Н	VH	3	3	3	3
Cascade Spring										
Upper Cowlitz <sup>c,g</sup>	Primary	VL	L	М	$VL^2$	H+	>500%	22,000	300	1,800
Cispus <sup>c,G</sup>	Primary	VL	L	M	VL <sup>2</sup>	H+	>500%	7,800	150	1,800
Tilton	Stabilizing	VL	VL	VL	VL	VL	0%	5,400	<100	
Toutle	Contributing	VL	Н	L	VL	М	>500%	3,100	100	1,100
Kalama	Contributing <sup>2</sup>	VL	Н	L	VL	L	>500%	4,900	100	300
Lewis NF <sup>c</sup>	Primary	VL	L	М	VL	Н	>500%	15,700	300	1,500
Sandy (OR) <sup>c,G</sup>	Primary	3	3	3	М	н	3	_3	3	3
Gorge Fall										
L. Gorge (WA/OR)	Contributing	VL	М	L	$VL^2$	М	>500%	n/a	<50	1,200
U. Gorge (WA/OR) C	Contributing <sup>1</sup>	VL	М	L	$VL^2$	M	>500%	n/a	<50	1,200
White Salmon c	Contributing	VL	L	L	VL	М	>500%	n/a	<50	500
Hood (OR)	Primary <sup>4</sup>	3	3	3	VL	н	3	3	3	3
Gorge Spring										
White Salmon <sup>c</sup>	Contributing	VL	VL	VL	VL	L+	>500%	n/a	<50	500
Hood (OR)	Primary	3	3	3	VL	VH	3	3	3	3
Source: LCERR 2010	•									

Source: LCFRB 2010.

L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.

I Increase relative to interim Plan.

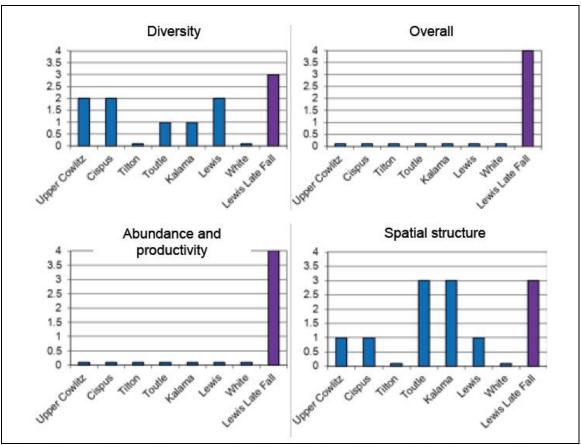
Reduction relative to interim Plan.

Reduction relative to interim Plan.

Addressed in Oregon Management Unit plan.

Designated as a historical core population by the TRT.

Designated as a historical legacy population by the TRT.



**Figure 2.2.2.1:** Current status of Washington lower Columbia River spring Chinook and late fall-run (bright) Chinook salmon populations for the VSP parameters and overall population risk. (LCFRB Recovery Plan 2010, chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

**Lower Columbia River Steelhead** (*Oncorhynchus mykiss*): The DPS includes all naturally spawned anadromous *O. mykiss* (steelhead) populations below natural and manmade impassable barriers in streams and tributaries to the Columbia River between the Cowlitz and Wind Rivers, Washington (inclusive), and the Willamette and Hood Rivers, Oregon (inclusive), and excludes fish originating from the upper Willamette River Basin above Willamette Falls. The DPS includes seven artificial propagation programs, including the Cowlitz Trout Hatchery Winter-late (Lower Cowlitz), Kalama River Wild (winter- and summer-run) and Lewis River Wild Winter (NMFS 2014 79FR20802).

**Status:** Today, 16 of the 23 Lower Columbia River steelhead populations have a low or very low probability of persisting over the next 100 years, and six populations have a moderate probability of persistence. Only the summer-run Wind population is considered viable. All four strata in the DPS fall short of the WLC TRT criteria for viability (Dornbush and Sihler 2013). Populations in the upper Lewis and Cowlitz watersheds remain cut-off from access to essential spawning habitat by hydroelectric dams. Projects to allow access have been initiated in the Cowlitz and Lewis systems but these have not yet produced self-sustaining populations (Ford 2011). Condit Dam on the White Salmon River was breached October 26, 2011. WDFW is currently developing watershed-specific management plans in accordance with the SSMP. As part of this planning process, WDFW is proposing to complete a thorough review of current steelhead stock status using the most up to date estimates of adult abundance, juvenile production and genetic information.

Table 2.2.2.2: Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River steelhead populations.

		Ва	seline	viabili	ty		Prod.	А	bundance	
Population	Contribution	A&P	S	D	Net	Obj.	target	Historical	Baseline	Target
Coast Winter										
Grays/Chinook	Primary	VH	VH	M	$M^1$	Н	0%4	1,600	800	800
Eloch/Skam	Contributing	VH	VH	M	$M^1$	M+	0%4	1,100	600	600
Mill/Ab/Germ	Primary	Н	VH	M	$M^1$	Н	0%4	900	500	500
Youngs Bay (OR)	Primary	3	3	3	VH	VH	3	3	3	3
Big Creek (OR)	Primary	3	3	3	Н	VH	3	3	3	3
Clatskanie (OR)	Primary	3	3	3	VH	VH	3	3	3	3
Scappoose (OR)	Primary	3	3	3	VH	VH	3	3	3	3
Cascade Winter										
Lower Cowlitz	Contributing	L	M	M	L	M	+5%	1,400	350	400
Upper Cowlitz <sup>c,g</sup>	Primary	VL	M	M	$VL^2$	$H^1$	>500%	1,400	<50	500
Cispus <sup>c,G</sup>	Primary	VL	M	M	$VL^2$	$H^1$	>500%	1,500	<50	500
Tilton	Contributing	VL	M	M	VL	L	>500%	1,700	<50	200
S.F. Toutle	Primary	M	VH	Н	M	H+	+35%	2.000	350	600
N.F. Toutle <sup>c</sup>	Primary	VL	Н	Н	$VL^2$	Н	+125%	3,600	120	600
Coweeman	Primary	L	VH	VH	L <sup>2</sup>	Н	+25%	900	350	500
Kalama	Primary	L	VH	Н	L <sup>2</sup>	H+	+45%	800	300	600
N.F. Lewis <sup>c</sup>	Contributing	VL	M	M	$VL^2$	M	>500%	8,300	150	400
E.F. Lewis	Primary	M	VH	M	$M^1$	Н	+25%	900	350	500
Salmon	Stabilizing	VL	Н	M	$VL^2$	VL	0%	na	<50	
Washougal	Contributing	L	VH	M	L <sup>2</sup>	M	+15%	800	300	350
Clackamas (OR) <sup>c</sup>	Primary	3	3	3	M	Н	3	3	3	3
Sandy (OR) <sup>c</sup>	Primary	3	3	3	L	VH	3	3	3	3
Cascade Summer										
Kalama <sup>c</sup>	Primary	Н	VH	M	$M^1$	Н	0%4	1,000	500	500
N.F. Lewis	Stabilizing	VL	VL	VL	VL	VL	0%	na	150	
E.F. Lewis <sup>G</sup>	Primary	VL	VH	M	$VL^2$	Н	>500%	600	<50	500
Washougal <sup>c,g</sup>	Primary	M	VH	M	$M^1$	Н	+40%	2,200	400	500
Gorge Winter										
L. Gorge (WA/OR)	Primary	L	VH	M	L <sup>2</sup>	Н	+45%	na	200	300
U. Gorge (WA/OR)	Stabilizing	L	M	M	L <sup>2</sup>	L	0%	na	200	
Hood (OR) <sup>c,g</sup>	Primary	_3	3	3	M	Н	3	3	3	3
Gorge Summer										
Wind <sup>c</sup>	Primary	VH	VH	Н	$H^1$	VH	0%4	na	1,000	1,000
Hood (OR)	Primary	_3	3	3	VL	Н	3	3	3	3

Source: LCFRB 2010.

L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.

<sup>&</sup>lt;sup>1</sup> Increase relative to interim Plan.

<sup>&</sup>lt;sup>2</sup> Reduction relative to interim Plan.

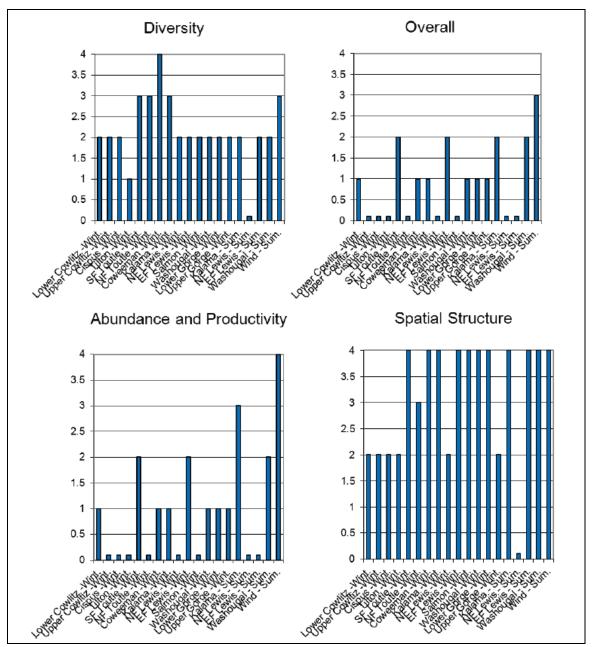
Addressed in Oregon Management Unit plan.

Management Unit plan.

Improvement in require improvement in spatial structure or diversity to meet recovery objectives.

C Designated as a historical core population by the TRT.

G Designated as a historical legacy population by the TRT.



**Figure 2.2.2.2:** Current status of Washington LCR steelhead populations for the VSP parameters and overall population risk. (LCFRB 2010 Recovery Plan, chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

Lower Columbia River coho (*Oncorhynchus kisutch*): Originally part of a larger Lower Columbia River/Southwest Washington ESU, Lower Columbia coho were identified as a separate ESU and listed as threatened on June 28, 2005. The ESU includes all naturally spawned populations of coho salmon in the Columbia River and its tributaries in Washington and Oregon, from the mouth of the Columbia up to and including the Big White Salmon and Hood Rivers, The twenty-one artificial propagation programs include: the Grays River, Peterson Coho Project, Cowlitz Type-N Coho Program in the Upper and Lower Cowlitz Rivers, Cowlitz Game and Anglers Coho Program, Friends of the Cowlitz Coho Program, North Fork Toutle River Hatchery, Kalama River Type-N and Type-S Coho Programs, Lewis River Type-N and Type-S

Coho programs, Fish First Wild Coho and Type-N Coho programs, Syverson Project Type-N Coho Program, and Washougal Hatchery Type-N Coho Program (NMFS 2014 79FR20802).

Status: Status evaluations of LCR coho status, all based on WLC-TRT criteria, have been conducted since the last BRT status update in 2005 (McElhany et al. 2007, Beamesderfer et al. 2010, LCFRB 2010, Dornbusch and Sihler 2013). All of these evaluations concluded that the ESU is currently at very high risk of extinction. All of the Washington side populations are considered at very high risk, although uncertainty is high because of a lack of adult spawner surveys. The 2005 BRT evaluation noted that smolt traps indicate some natural production in Washington populations, though given the high fraction of hatchery origin spawners suspected to occur in these populations it is not clear that any are self-sustaining (Ford 2011). Since this time WDFW has implemented an ESU wide monitoring program for LCR coho which began in 2010. Preliminary results indicate that natural origin population abundance may be higher than previously thought for certain populations (WDFW, unpublished). Results from the first 3 years of monitoring should be available in the near future. Currently, 21 of the 24 Lower Columbia River coho salmon populations are considered to have a very low probability of persisting over the next 100 years, and none is considered viable (Dornbusch and Sihler 2013). All three strata in the ESU fall significantly short of the WLC TRT criteria for viability.

**Table 2.2.2.3:** Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River coho populations.

		Ba	seline	viabil	ity		Prod.	Д	Abundance	
Population	Contribution	A&P	s	D	Net	Obj.	target	Historical	Baseline	Target
<u>Coast</u>										
Grays/Chinook L	Primary	VL	Н	VL	$VL^2$	Н	+370%	3,800	<50	2,400
Eloch/Skam L	Primary	VL	Н	VL	$VL^2$	Н	+170%	6,500	<50	2,400
Mill/Ab/Germ L	Contributing	VL	Н	L	$VL^2$	M	>500%	2,800	<50	1,800
Youngs (OR) L	Stabilizing	3	3	3	VL	VL	3	3	3	3
Big Creek (OR)	Stabilizing <sup>2</sup>	3	3	3	VL	VL	3	3	3	3
Clatskanie (OR) L	Primary <sup>1</sup>	3	3	3	L	VH	3	3	3	3
Scappoose (OR) L	Primary	3	3	3	М	VH	3	3	3	3
<u>Cascade</u>										
Lower Cowlitz L	Primary	VL	M	M	VL <sup>2</sup>	Н	+100%	18,000	500	3,700
Upper Cowlitz <sup>E, L</sup>	Primary <sup>1</sup>	VL	M	L	VL	H1	>500%	18,000	<50	2,000
Cispus <sup>E, L</sup>	Primary <sup>1</sup>	VL	M	L	VL	H1	>500%	8,000	<50	2,000
Tilton <sup>E, L</sup>	Stabilizing <sup>2</sup>	VL	M	L	VL	VL <sup>2</sup>	0%	5,600	<50	
Toutle SF E, L	Primary	VL	Н	M	VL <sup>2</sup>	Н	+180%	27,000	<50	1,900
Toutle NF E, L	Primary	VL	M	L	$VL^2$	Н	+180%	27,000	<50	1,900
Coweeman L	Primary	VL	Н	M	VL <sup>2</sup>	Н	+170%	5,000	<50	1,200
Kalama <sup>L</sup>	Contributing	VL	Н	L	$VL^2$	L	>500%	800	<50	500
NF Lewis <sup>E, L</sup>	Contributing	VL	L	L	$VL^2$	L	+50%	40,000	200	500
EF Lewis <sup>E, L</sup>	Primary	VL	Н	M	$VL^2$	Н	>500%	3,000	<50	2,000
Salmon L	Stabilizing	VL	M	VL	VL	VL	0%	na	<50	
Washougal L	Contributing	VL	Н	L	VL <sup>2</sup>	M+	>500%	3,000	<50	1,500
Clackamas (OR) E, L	Primary	_3	3	3	M	VH	3	_3	3	3
Sandy (OR) E, L	Primary	3	3	3	VL	Н	3	3	3	3
Gorge										
L Gorge (WA/OR) L	Primary	VL	M	VL	$VL^2$	Н	+400%	na	<50	1,900
U Gorge (WA) L	Primary <sup>1</sup>	VL	M	VL	VL <sup>2</sup>	Н	+400%	na	<50	1,900
U Gorge/Hood (OR) <sup>E</sup>	Contributing <sup>4</sup>	3	3	3	VL	Н	3	3	3	3

Source: LCFRB 2010.

L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.

(Core and Legacy populations not designated by the TRT for coho).

<sup>&</sup>lt;sup>1</sup> Increase relative to interim Plan.

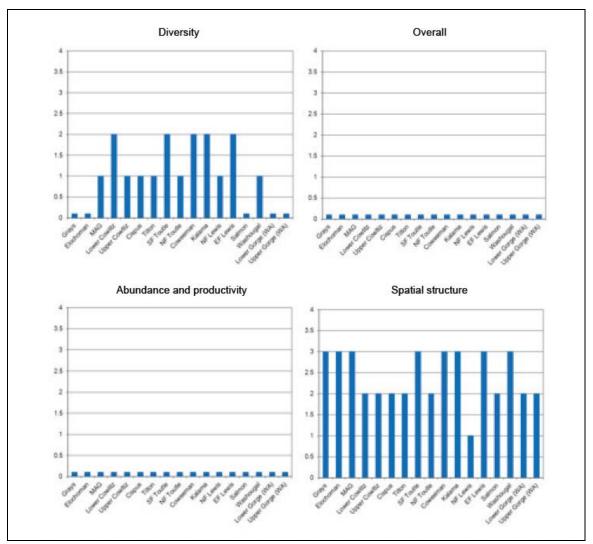
<sup>&</sup>lt;sup>2</sup> Reduction relative to interim Plan.

<sup>&</sup>lt;sup>3</sup> Addressed in Oregon Management Unit plan.

<sup>&</sup>lt;sup>4</sup> Improvement increments are based on abundance and productivity; however, this population will require improvement in spatial structure or diversity to meet recovery objectives.

<sup>&</sup>lt;sup>E</sup> Early run (Type S) coho stock.

<sup>&</sup>lt;sup>L</sup> Late run (Type N) coho stock.



**Figure 2.2.2.3:** Current status of Washington LCR coho populations for the VSP parameters and overall population risk. (LCFRB 2010 recovery plan, chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

**Columbia River chum salmon** (*Oncorhynchus keta*). ESU includes all naturally spawned populations of chum salmon in the Columbia River and its tributaries in Washington and Oregon, as well as artificial propagation programs at Grays River and Washougal River/Duncan Creek chum hatchery programs (NMFS 2014 79FR20802).

**Status:** A report on the population structure of lower Columbia River salmon and steelhead populations was published by the WLC-TRT in 2006 (Myers et al. 2006). The chum population designations in that report are used in this status update and were used for status evaluations in recent recovery plans by ODFW and LCFRB.

The LCFRB completed a revision recovery plan in 2010 that includes Washington populations of Columbia River chum salmon. This plan includes an assessment of the current status of Columbia River chum populations, which relied and built on the viability criteria developed by the WLC-TRT (McElhany et al. 2006) and an earlier evaluation of Oregon WLC populations (McElhany et al. 2007). This evaluation assessed the status of populations with regard to the VSP parameters of A/P, spatial structure, and diversity (McElhany et al. 2000). The result of this analysis is shown in **Figure 2.2.2.3**. The analysis indicates that all of the Washington populations with two exceptions

are in the overall very high risk category (also described as extirpated or nearly so). The Grays River population was considered to be at moderate risk and the Lower Gorge population to be at low risk. The very high risk status assigned to the majority of Washington populations (and all the Oregon populations) reflects the very low abundance observed in these populations (e.g., <10 fish/year) (Ford 2011). Today, 15 of the 17 populations that historically made up this ESU are so depleted that either their baseline probability of persistence is very low or they are extirpated or nearly so; this is the case for all six of the Oregon populations. Currently almost all natural production occurs in just two populations: Grays/Chinook and the Lower Gorge. All three strata in the ESU fall significantly short of the WLC TRT criteria for viability (Dornbush and Sihler 2013).

**Table 2.2.2.4:** Baseline viability status, viability and abundance objectives, and productivity improvement targets for lower Columbia River chum populations.

		Ва	seline	viabili	ty		Prod.	Abundance		
Population	Contribution	A&P	S	D	Net	Obj.	target	Historical	Baseline	Target
Coast										
Grays/Chinook <sup>c,G</sup>	Primary	VH	M	Н	$M^1$	VH	0%4	10,000	1,600	1,600
Eloch/Skam <sup>c</sup>	Primary	VL	Н	L	$VL^2$	Н	>500%	16,000	<200	1,300
Mill/Ab/Germ	Primary	VL	Н	L	VL	Н	>500%	7,000	<100	1,300
Youngs (OR) <sup>c</sup>	Stabilizing <sup>2</sup>	3	3	3	VL	VL	3	3	3	3
Big Creek (OR) <sup>C</sup>	Stabilizing <sup>2</sup>	3	3	3	VL	VL	3	3	3	3
Clatskanie (OR)	Primary <sup>1</sup>	3	3	3	VL	Н	3	3	3	3
Scappoose (OR)	Primary <sup>1</sup>	3	3	3	VL	Н	3	3	3	3
<u>Cascade</u>										
Cowlitz (Fall) <sup>c</sup>	Contributing	VL	Н	L	VL	M	>500%	195,000	<300	900
Cowlitz (Summer) <sup>c</sup>	Contributing	VL	L	L	VL	M	>500%	n/a	n/a	900
Kalama	Contributing	VL	Н	L	VL	M	>500%	20,000	<100	900
Lewis <sup>c</sup>	Primary	VL	Н	L	VL	Н	>500%	125,000	<100	1,300
Salmon	Stabilizing	VL	L	L	VL	VL	0%	n/a	<100	
Washougal	Primary	VL	Н	L	$VL^2$	H+	>500%	18,000	<100	1,300
Clackamas (OR) <sup>c</sup>	Contributing	3	3	3	VL	M	3	3	3	3
Sandy (OR)	Primary	3	3	3	VL	Н	3	3	3	3
Gorge										
L. Gorge (WA/OR) C,G	Primary	VH	Н	VH	$H^1$	VH	0%4	6,000	2,000	2,000
U. Gorge (WA/OR)	Contributing	VL	L	L	VL	M	>500%	11,000	<50	900

Source: LCFRB 2010.

L = Low; M = Moderate; H = High; VH/E = Very High/Extinct.

<sup>&</sup>lt;sup>5</sup> Increase relative to interim Plan.

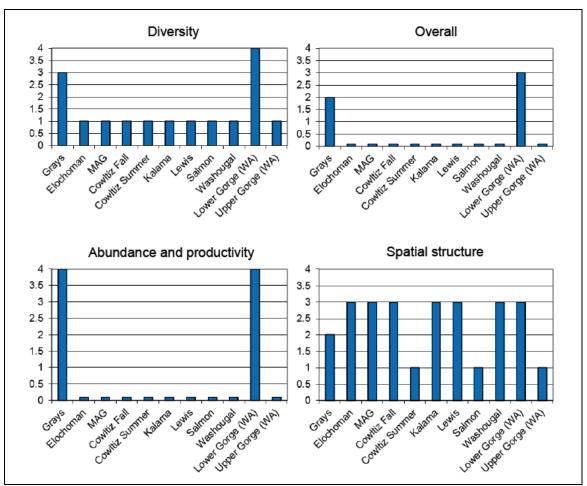
<sup>&</sup>lt;sup>6</sup> Reduction relative to interim Plan.

<sup>&</sup>lt;sup>7</sup> Addressed in Oregon Management Unit plan.

<sup>8</sup> Improvement increments are based on abundance and productivity; however, this population will require improvement in spatial structure or diversity to meet recovery objectives.

<sup>&</sup>lt;sup>C</sup> Designated as a historical core population by the TRT.

<sup>&</sup>lt;sup>G</sup> Designated as a historical legacy population by the TRT.



**Figure 2.2.2.4:** Current status of Washington CR chum populations for the VSP parameters and overall population risk. (LCFRB 2010 Recovery Plan, Chapter 6). A population score of zero indicates a population extirpated or nearly so, a score of 1 is high risk, 2 is moderate risk, 3 is low risk ("viable") and 4 is very low risk (Ford 2011).

## - Provide the most recent 12 year (e.g. 1988-present) progeny-to-parent ratios, survival data by life-stage, or other measures of productivity for the listed population.

Not available for most species. See HGMP section 11.1 for planned M&E. Juvenile coho production estimates is the one measure of production in the Lower Columbia system.

**Table 2.2.2.5:** Lower Columbia River Washington tributary coho smolt production estimates, 1997-2009 (WDFW, Region 5).

Year	Cedar Creek	Mill Creek	Abernathy Creek	Germany Creek	Cowlitz Falls Dam	Mayfield Dam
1997					3,700	700
1998	38,400				110,000	16,700
1999	28,000				15,100	9,700
2000	20,300				106,900	23,500
2001	24,200	6,300	6,500	8,200	334,700	82,200
2002	35,000	8,200	5,400	4,300	166,800	11,900
2003	36,700	10,500	9,600	6,200	403,600	38,900
2004	37,000	5,700	6,400	5,100	396,200	36,100
2005	58,300	11,400	9,000	4,900	766,100	40,900

2006	46,000	6,700	4,400	2,300	370,000	33,600
2007	29,300	7,000	3,300	2,300	277,400	34,200
2008	36,340	90,97	5,077	3,976		38,917
2009	61,140	62,83	3,761	2,576		29,718
2010						49,171
2011						43,831

Source: LCR FMEP Annual Report 2010 and WDFW Data 2012.

## - Provide the most recent 12 year annual spawning abundance estimates, or any other abundance information. Indicate the source of these data.

**Table 2.2.2.6:** Spring Chinook salmon total spawner abundance estimates in LCR tributaries, 2000-2012.

Year	Cowlitz	Kalama	Lewis
2000	266	34	523
2001	347	578	754
2002	419	898	498
2003	1,953	790	745
2004	1,877	358	529
2005	405	380	122
2006	783	292	857
2007	74	2,150	264
2008	425	364	40
2009	763	34	80
2010	711	0	160
2011	1,359	26	120
2012	1,359	28	200

Source: Joe Hymer, WDFW Annual Database 2012

**Table 2.2.2.7:** Fall Chinook salmon total spawner abundance estimates in LCR tributaries, 2000-2011<sup>a</sup>.

2011.											
Year	Elochoman River	Coweeman River <sup>a</sup>	Grays River	Skamokaw a Creek	Cowlitz River	Green River (Toutle)	SF Toude River	Kalama River	EF Lewis River	NF Lewis River	Washougal River
2000	884	424	80	482	2,100	1,580	204	3,877	391	6,504	2,757
2001	230	251	104	3	1,979	1,081	102	3,451	245	4,281	1,704
2002	332	566	390	7	3,038	5,654	216	10,560	441	5,518	2,728
2003	2,204	753	149	529	2,968	2,985	327	9,272	607	11,519	2,678
2004	4,796	1,590	745	2,109	4,621	4,188	618	6,680	918	13,987	10,597
2005	6,820	1,090	387	588	10,329	13,846	140	24,782	727	18,913	3,444
2006	7,581	900	82	372	14,427	7,477	450	18,952	1,375	17,106	6,050
2007	194	140	99	36	2,724	961	30	1,521	308	10,934	2,143
2008	782	95	311	253	1,334	824	45	2,617	236	4,268	3,182
2009	231	147	93	139	2,156	1,302	66	4,356	110	6,112	2,995
2010	1,883	1,330	12	268	2,762	605	NE	3,576	314	8,908	4,529
2011	508	2,148	353	41	1,616	668	NE	10,639	334	14,033	2,961
	D D 1	TT I D D TT I	NT . 10	ъ.		. 2012					

Source: Ron Roler, WDFW Natural Spawn Progress Reports 2012.

<sup>\*</sup> Estimates of total adult and jack fall Chinook. May include fish put upstream of hatchery weirs.

Table 2.2.2.8: Wild winter steelhead escapement estimates for select SW Washington DPS

populations, current WDFW escapement goals and LCSRP abundance targets.

Location	Grays River	Elochoman/ Skamokawa	Mill/Abernathy/ Germany
WDFW			
<b>Escapement Goal</b>	1,486	853	508
LCSRP			
Abundance Target	800	600	500
2000	1,064	650	380
2001	1,130	656	458
2002	724	370	354
2003	1,200	668	342
2004	1,132	768	446
2005	396	376	274
2006	718	632	398
2007	724	490	376
2008	764	666	528
2009	568	222	396
2010	422	534	398
2011	318	442	270
3-year average	436	399	355
5-year average	559	471	394
10-year average	697	517	378

Source: WDFW Data 2012

 Table 2.2.2.9: Wild winter steelhead escapement estimates for select SW Washington DPS

populations, current WDFW escapement goals and LCSRP abundance targets.

populations, earrent			NF Toutle/		8	
Location	Coweeman	SF Toutle	Green	Kalama	EF Lewis	Washougal
WDFW						
<b>Escapement Goal</b>	1,064	1,058	NA	1,000	1,243	520
LCSRP						
Abundance Target	500	600	600	600	500	350
2000	530	490		921	NA	NA
2001	384	348		1,042	377	216
2002	298	640		1,495	292	286
2003	460	1,510		1,815	532	764
2004	722	1,212		2,400	1,298	1,114
2005	370	520	388	1,856	246	320
2006	372	656	892	1,724	458	524
2007	384	548	565	1,050	448	632
2008	722	412	650	776	548	732
2009	602	498	699	1,044	688	418
2010	528	274	508	961	336	232
2011	408	210	416	622	308	204
3-year average	513	327	541	876	444	285
5-year average	529	388	568	891	466	444
10-year average	487	648	*588	1,374	515	523

Source: WDFW Data 2012.

<sup>\* 7-</sup>year average for NF Toutle/Green.

Table 2.2.2.10: Wild summer steelhead population estimates for LCR populations from 2001 to

2011, current WDFW escapement goals, and LCSRP abundance targets.

Location	Kalama	EF Lewis	Washougal	Wind
WDFW Escapement Goal	1,000	NA	NA	1,557
LCSRP Abundance Target	500	500	500	1,000
2001	286	271	184	457
2002	454	440	404	680
2003	817	910	607	1,096
2004	632	425	NA	861
2005	400	673	608	587
2006	387	560	636	632
2007	361	412	681	737
2008	237	365	755	614
2009	308	800	433	580
2010	370	602	787	788
2011	534	1,084*	956*	1,468
3-year average	404	829	725	945
5-year average	362	653	722	837
10-year average	450	627	652	804

Source: WDFW Data 2012. \* Preliminary estimates.

**Table 2.2.2.11:** Population estimates of chum salmon in the Columbia River.

Location	2002	2003	2004	2005	2006	2007	2008	2009	2010 <sup>a</sup>	2011 <sup>a</sup>
Crazy Johnson Creek			966	1,471	3,639	759	1,034	981	677	2,374
WF Grays River			9,015	1,324	1,232	1,909	800	994	1,967	7,002
Mainstem Grays River			4,872	1,400	1,244	1,164	886	750	3,467	1,848
I-205 area	3,468	2,844	2,102	1,009	862	544	626	1,132	2,105	4,947
Multnomah area	1,267	1,130	665	211	313	115	28	102	427	641
St Cloud area		137	104	92	173	9	1	14	99	509
Horsetail area			106	40	63	17	33	6	45	183
Ives area <sup>b</sup>	4,466	1,942	363	263	387	145	168	141	214	162
Duncan Creek <sup>c</sup>	13	16	2	7	42	9	2	26	48	85
Hardy Creek	343	392	49	73	104	14	3	39	137	173
Hamilton Creek	1,000	500	222	174	246	79	114	115	247	517
Hamilton Spring Channel	794	363	346	84	236	44	109	91	187	324
Grays return <sup>d</sup>	12,041	16,974	15,157	4,327	6,232	3,966	2,807	2,833	6,399	11,518
I-205 to Bonneville return	11,351	7,324	3,959	1,953	2,426	976	1,084	1,666	3,509	7,541
Lower Columbia River Total	23,392	24,298	19,116	6,280	8,658	4,942	3,891	4,499	9,908	19,059

Source: Todd Hillson - WDFW Chum Program 2012

<sup>&</sup>lt;sup>a</sup> Data for 2010 and 2011 is preliminary.

b Ives area counts are the carcass tagging estimate plus fish removed for broodstock, except for 2007 and 2008, which is area under the curve.

<sup>&</sup>lt;sup>c</sup> Totals for Duncan Creek do not include broodstock brought in from mainstem spawning areas, adult trap catch or surveys below monitoring weirs only..

<sup>&</sup>lt;sup>d</sup> Grays return totals include natural spawners and removed for broodstock.

- Provide the most recent 12 year (e.g. 1988-1999) estimates of annual proportions of direct hatchery-origin and listed natural-origin fish on natural spawning grounds, if known.

The proportion of effective hatchery-origin spawners (pHOS) should be less than 30% of the naturally spawning population for this integrated program per HSRG guidelines (2009), as it is associated with a Primary natural population. Current pHOS estimates are 0.51. See HGMP section 11.1 for planned M&E.

- 2.2.3 <u>Describe hatchery activities, including associated monitoring and evaluation and research programs, that may lead to the take of NMFS listed fish in the target area, and provide estimated annual levels of take.</u>
- Describe hatchery activities that may lead to the take of listed salmonid populations in the target area, including how, where, and when the takes may occur, the risk potential for their occurrence, and the likely effects of the take.

#### **Broodstock Program:**

*Broodstock Collection:* In previous years, hatchery Chinook were not mass-marked, and the level of listed Chinook encountered during coho collection was unknown. Listed fish above broodstock needs and identified by the presence of the adipose fin are released upstream although trapping operations cease by mid to late fall, prior to the wild winter steelhead run. See "take" tables at the end of this document.

Genetic introgression: Broodstock for this program was initiated from local coho salmon. The stock was managed to maintain characteristics of the Type S Coho since the 1950s (see HGMP section 1.13). Egg-takes are representative of adult arriving throughout the run and the current collection protocol preserves the range timing of the historical coho stock in the system. Most natural spawners in the system are composites and representative of the lower Columbia coho (SaSI 2002). Large numbers of hatchery spawners are released upstream. There is approximately ¼-mile of spawning area below the hatchery weir (between the weir and the confluence with the NF Toutle).

#### **Rearing Program:**

Operation of Hatchery Facilities: Facility operation impacts include water withdrawal, effluent, and intake compliance. Effluent at outfall areas is rapidly diluted with mainstem flows and operation is within permitted NPDES guidelines (see HGMP sections 4.1 and 4.2). Indirect take from this operation is unknown.

Disease: Over the years, rearing densities, disease prevention and fish health monitoring have greatly improved the health of the hatchery programs. *Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries*-Chapter 5 (IHOT 1995) have been instrumental in reducing disease outbreaks. Although pathogens occur in the wild and fish might be affected, they are believed to go undetected with predation quickly removing those fish.

In addition, although pathogens may cause post release mortality in fish from hatcheries, there is little evidence that hatchery origin fish routinely infect natural populations of salmon and steelhead in the Pacific Northwest (Enhancement Planning Team 1986 and Steward and Bjornn 1990). Prior to release, the hatchery population health and condition is established by the Area Fish Health Specialist. This is commonly done one to three weeks pre-release, and up to six weeks on systems with pathogen-free water and little or no history of disease. Indirect take from disease is unknown.

#### Release:

Hatchery Production/Density-Dependent Effects: Hatcheries can release numbers of fish that can exceed the density of the natural productivity in a limited area for a short period of time and can

compete with listed fish. Fish are released as active smolts that will emigrate in order to minimize the effect of the release. Indirect take from density dependent effects is unknown.

Potential Lewis coho predation and competition effects on listed salmonids and eulachon: The proposed annual production goal for this program is 150,000 yearlings. Coho are released at 15 fpp (146 mm fl). Due to size differences between coho smolts and fingerling listed stocks, competition is unlikely with different prey items and habitat preferences. Indirect take from predation is unknown.

**Table 2.2.3.1:** Peak migration timing and average fork length (mm) of out-migrant juvenile Chinook, coho and steelhead captured in rotary screw traps on Mill, Germany and Abernathy creek, Lower Columbia River, 2008.

	(	Chinook		Coho	Steelhead		
Stream	Avg Size (mm)	Peak Migration	Avg Size (mm)	Peak Migration	Avg Size (mm)	Peak Migration	
Mill Cr	37.0	Mar 10-Apr 13	104.2	Mar 17-23	154.5	Apr 28-May 4	
Germany Cr	39.8	Mar 17-23	115.3	May 19-25	177.8	May 12-18	
Abernathy Cr	37.9	Mar 31 – Apr 6	112.1	May 19-25	163.8	May 12-18	

Source: Kinsel et al 2009.

Both juvenile and adult salmonids have been documented to feed on eulachon (Gustafson et al. 2010). Predation of eulachon by steelhead reared in this program may occur, however it is unknown to what degree such predation may occur.

*Residualism*: To maximize smolting characteristics and minimize residualism, WDFW adheres to a combination of acclimation, volitional release strategies, size, and time guidelines.

- Condition factors, standard deviation and co-efficient of variation (CV) are measured throughout the rearing cycle and at release.
- Feeding rates and regimes throughout the rearing cycle are programmed to satiation feeding to minimize out-of-size fish and programmed to produce smolt size fish at date of release.
- Based on past history, fish have reached a size and condition that indicates a smolted condition at release.
- Releases occur within known time periods of species emigration from acclimated ponds.
- Releases from these ponds are volitional with large proportions of the populations moving out initially with the remainder of the population vacating within days or a few weeks.

#### **Monitoring:**

Associated monitoring Activities: Interaction between hatchery and wild adult salmonids will be managed by monitoring key tributary escapements of coho, steelhead, cutthroat and chum.

The following monitoring baseline activities are conducted in the Lower Columbia Management Area (LCMA) for adult steelhead and salmon: redd surveys are conducted for fall Chinook in the SF Toutle, Coweeman, EF Lewis and Washougal rivers. Redd surveys are also conducted in the Cowlitz River for fall and spring Chinook. Mark-recapture surveys provide data for summer steelhead populations in the Wind and Kalama rivers. Mark-recapture carcass surveys are conducted to estimate populations of Chinook salmon in Grays, Elochoman, Coweeman, SF Toutle, Green, Kalama, NF Lewis, EF Lewis, rivers and Skamokawa, Mill, Abernathy, and Germany creeks and for all chum salmon populations. Snorkel surveys are conducted for summer steelhead in the EF Lewis, Washougal Rivers. Trap counts are conducted on the Cowlitz, NF Toutle, Kalama, and Wind rivers and on Cedar Creek a tributary of the NF Lewis River. Area-Under-the-Curve (AUC) surveys are conducted to collect population data for chum salmon in Grays River and Hardy and Hamilton Creeks. All sampling of carcasses and trapped fish include recovery of coded wide tagged (CWT) fish for hatchery or wild stock evaluation. Downstream migrant trapping occurs on the Cowlitz, Kalama, NF Lewis, and Wind rivers, Cedar Creek, and

will expand to other basins as part of a salmonid life cycle monitoring program to estimate freshwater production and wild smolt to adult survival rates. Any take associated with monitoring activities is unknown but all follow scientific protocols designed to minimize impact.

- Provide information regarding past takes associated with the hatchery program, (if known) including numbers taken, and observed injury or mortality levels for listed fish.

**Table 2.2.3.2:** Disposition of unmarked (no adipose fin-clip) coho returning to North Toutle Hatchery.

Brood Year	Mortality	Spawned
2004	0	0
2005	0	0
2006	1	18
2007	6	97
2008	11	20
2009	29	76
2010	14	52
2011	17	129
2012	23	144
2013	20	116
Average		

Source: WDFW Annual Escapement Reports.

- Provide projected annual take levels for listed fish by life stage (juvenile and adult) quantified (to the extent feasible) by the type of take resulting from the hatchery program (e.g. capture, handling, tagging, injury, or lethal take).

See "take" tables at the end of this HGMP.

- Indicate contingency plans for addressing situations where take levels within a given year have exceeded, or are projected to exceed, take levels described in this plan for the program.

No situations are expected to occur where take would exceed ESA limits. If significant numbers of wild salmonids are observed impacted by this operation, then staff would inform the WDFW District Biologist, Fish Health Specialist or Area Habitat Biologist who, along with the Hatchery Complex Manager, would determine an appropriate plan and consult with NOAA-NMFS for adaptive management review and protocols.

Handling and release of wild coho in broodstock trapping operations is monitored and take observations have been rare. Any additionally mortality from this operation on a yearly basis would be communicated to Fish program staff for additional guidance.

#### 3 <u>SECTION 3. RELATIONSHIP OF PROGRAM TO OTHER</u> MANAGEMENT OBJECTIVES

3.1 Describe alignment of the hatchery program with any ESU-wide hatchery plan (e.g. Hood Canal Summer Chum Conservation Initiative) or other regionally accepted policies (e.g. the NPPC Annual Production Review Report and Recommendations - NPPC document 99-15). Explain any proposed deviations from the plan or policies.

WDFW has several policies/plans that help inform management decisions regarding the HGMPs currently under review. These policies include:

- 1. Hatchery and Fishery Reform Policy (Commission Policy C3619)
- 2. The Conservation and Sustainable Fisheries Plan (draft)
- 3. The Hatchery Action Implementation Plans (HAIP)
- 4. Lower Columbia Salmon Recovery Plan (LCSRP)

Descriptions of these policies and excerpts are shown below:

#### Policies/Plans – Key Excerpts

Hatchery and Fishery Reform Policy: Washington Department of Fish and Wildlife Commission Policy C-3619. WDFW adopted the Hatchery and Fishery Reform Policy C-3619 in 2009. Its purpose is to advance the conservation and recovery of wild salmon and steelhead by promoting and guiding the implementation of hatchery reform. The intent of hatchery reform is to improve hatchery effectiveness, ensure compatibility between hatchery production and salmon recovery plans and rebuilding programs, and support sustainable fisheries. WDFW Policy C-3619 works to promote the conservation and recovery of wild salmon and steelhead and provide fishery-related benefits by establishing clear goals for each state hatchery, conducting scientifically defensible-operations, and using informed decision making to improve management. It is recognized that many state operated hatcheries are subject to provisions under U.S. v Washington (1974) and U.S. v Oregon and that hatchery reform actions must be done in close coordination with tribal comanagers. Washington Fish and Wildlife Commission Policy: POL-C3619.

Guidelines from the policy include:

- 1. Use the principles, standards, and recommendations of the Hatchery Scientific Review Group (HSRG) to guide the management of hatcheries operated by the Department.
- 2. Develop watershed-specific action plans that systematically implement hatchery reform as part of a comprehensive, integrated (All-H) strategy for meeting conservation and harvest goals at the watershed and Evolutionarily Significant Unit (ESU)/Distinct Population Segment (DPS) levels. Action Plans will include development of stock (watershed) specific population designations and application of HSRG broodstock management standards.

Conservation and Sustainable Fisheries Plan (CSFP): The CSFP is a draft plan that has been developed to meet WDFW's responsibilities outlined in the Lower Columbia Salmon Recovery Plan (LCSRP) and address the HSRG suggested solutions and achieve HRSG standards for primary, contributing and stabilizing populations. The plan describes the implementation of changes to hatchery and harvest programs and how they assist in recovery and achieve HSRG guidelines. The draft plan also identifies Viable Salmonid Population (VSP) parameters that will be addressed.

Hatchery Action Implementation Plans (HAIP): The HAIPs illustrate how WDFW is implementing hatchery programs to incorporate the HSRG guidelines. The plans provide the current programs and explain the future goals.

Lower Columbia Salmon Recovery Plan (LCSRP): Some sub-basins will be free of hatchery influence and hatchery programs. In other sub-basins, hatchery programs will serve specific conservation and harvest purposes consistent with goals for naturally-spawning populations. The mosaic of programs is designed to ensure that overall each DPS will be naturally self-sustaining.

#### **Strategies**

- 1. Reconfigure production-based hatchery programs to minimize impacts on natural populations and complement recovery objectives.
- 2. Adaptively manage hatcheries to respond to future knowledge, enhance natural production, and improve operational efficiencies.

Mitchell Act: This program receives Mitchell Act Funding. Initially passed in 1938, the Mitchell Act is intended to help rebuild and conserve the fish runs, and mitigate the impacts to fish from water diversions, dams on the mainstem of the Columbia River, pollution and logging. The Mitchell Act specifically directs establishment of salmon hatcheries, conduct of engineering and biological surveys and experiments, and installing fish protective devices. It also authorizes agreements with State fishery agencies and construction of facilities on State-owned lands. NMFS has administered the program as of 1970. There are 15 Mitchell Act hatcheries in Washington State; the majority of which are below Bonneville Dam.

The Mitchell Act programs are intended to support Northwest fishing economies – particularly coastal and Native American -- that have relied on Columbia River production both before and after dam construction. Catches of hatchery fish sustain the economies of local communities while keeping incidental mortalities of ESA-Listed fish at approved levels. Value of hatchery production and benefit to local economies will be further increased by implementing fisheries that increase harvest of hatchery produced fish, as expected through implementation of the LCSRP.

## 3.2 List all existing cooperative agreements, memoranda of understanding, memoranda of agreement, or other management plans or court orders under which program operates.

Future Brood Document. Hatchery salmon and steelhead production levels are detailed in the annual Future Brood Document, a pre-season planning document for fish hatchery production in Washington State for the upcoming brood stock collection and fish rearing season (July 1 – June 30).

See also HGMP section 3.1.

#### 3.3 Relationship to harvest objectives.

Total annual harvest is dependent on management response to annual abundance in Pacific Salmon Commission (PSC - U.S./Canada), Pacific Fishery Management Council (PFMC - U.S. ocean), and Columbia River Compact forums. WDFW also has received authorization for tributary, Columbia River mainstem, and ocean fisheries; the combined harvest rates in the *Fisheries Management and Evaluation Plan* (FMEP), *Columbia River Fish Management Plan* (CRFMP), and ocean fisheries are reviewed annually in the North of Falcon process.

## 3.3.1 <u>Describe fisheries benefitting from the program, and indicate harvest levels and rates for program-origin fish for the last twelve years (1988-99), if available.</u>

Hatchery coho can contribute significantly to the lower Columbia River gill net fishery; commercial harvest of early coho is constrained by fall Chinook and Sandy River coho management; commercial harvest of late coho is focused in October during the peak abundance of hatchery late coho. A substantial estuary sport fishery exists between Buoy 10 and the Astoria-Megler Bridge; majority of the catch is early hatchery coho, but late hatchery coho harvest can also be substantial.

**Table 3.3.1.1:** North Toutle Hatchery Type-S Coho fisheries contributions.

Brood Years: 2001-2005 Fishery Years: 2003-2008

	Average SAR%	2.39
Agency	Non-WA Fishery	% Total Survival
CDFG	All	0.7
Agency	Non-WA Fishery	% Total Survival
ODFW	10- Ocean Troll	1.1
ODFW	21- Columbia R. Gillnet	1.8
ODFW	40- Ocean Sport	10.3
ODFW	44- Columbia R. Sport	0.2
ODFW	45- Estuary Sport-(Buoy 10)	2.2
ODFW	72- Juvenile Sampling - Seine (Marine)	0.1
Agency	Non-WA Fishery	% Total Survival
WDFW	10- Ocean Troll	0.2
WDFW	15- Treaty Troll	0.6
WDFW	41- Ocean Sport- Charter	2.9
WDFW	42- Ocean Sport- Private	6.8
WDFW	45- Estuary Sport	0.1
WDFW	46- Freshwater Sport	3.6
Unk	50- Hatchery Escapement	0.0
WDFW	50- Hatchery Escapement	69.3
	Total	100.0

Source: RMIS 2012.

#### 3.4 Relationship to habitat protection and recovery strategies.

The following processes have included habitat identification problems, priority fixes and evolved as key components to *The Lower Columbia Salmon Recovery and Fish and Wildlife Sub-basin Plans* (Volume 1; Clark, Cowlitz, Lewis, Skamania and Wahkiakum Counties, December 15, 2004).

Sub-Basin Planning - Regional sub-basin planning processes include the Cowlitz River Sub-basin Salmon and Steelhead Production Plan, September 1, 1990 with a more recent Draft Cowlitz River Sub-basin Summary (May 17, 2002) was prepared for the Northwest Power Planning Council. The Sub-basin efforts provided initial building blocks for the LCFRB regional recovery plan. The Lower Columbia fish Recovery Board (LCFRB) has adopted the Lower Columbia Salmon Recovery and Fish and Wildlife Sub-basin Plans (Volume 1; Clark, Cowlitz, Lewis, Skamania and Wahkiakum Counties, December 15, 2004) with the understanding that Implementation of the schedule and actions for local jurisdictions depends upon funding and other resources.

Habitat Treatment and Protection - Ecosystem Diagnosis and Treatment (EDT) compares habitat today to that of the basin in a historically unmodified state. EDT has been modeled for productivity in the Cowlitz basin in The Lower Columbia Salmon Recovery and Fish and Wildlife Sub-basin Plans and has been used by Tacoma Power for the FERC re-licensing agreements for the upper basin productivity goals. WDFW is also conducting a Salmon Steelhead Habitat Inventory Assessment Program (SSHIAP), which documents barriers to fish passage. WDFW's habitat program issues hydraulic permits for construction or modifications to streams and wetlands. This provides habitat protection to riparian areas and actual watercourses within the watershed.

Average SAR% = (tags recovered/tags released)

<sup>\*</sup>Freshwater Sport based on WDFW Catch Record Card (CRC) data for BYs 2001-2005.

Limiting Factors Analysis (LFA) - A WRIA 26 LFA was conducted by the Washington State Conservation Commission (May 2002). WRIA 26 was separated into seven sub-basins; Coweeman, Lower Cowlitz, Toutle, Mayfield/Tilton, Riffe Lake, Cispus, and Upper Cowlitz.

#### 3.5 Ecological interactions.

- (1) Salmonid and non-salmonid fishes or species that could negatively impact the program: Outmigrant hatchery fish can be preyed upon through the entire migration corridor from the river sub-basin to the mainstem Columbia River and estuary. Northern pikeminnows and introduced spiny rays, as well as avian predators, including gulls, mergansers, cormorants, belted kingfishers, great blue herons and night herons in the Columbia mainstem sloughs, can prey on coho smolts. Mammals that can take a heavy toll on migrating smolts and returning adults include: harbor seals, sea lions, river otters and orcas
- (2) Salmonid and non-salmonid fishes or species that could be negatively impacted by the program: Co-occurring natural salmon and steelhead populations in local tributary areas and the Columbia River mainstem corridor areas could be negatively impacted by program fish. Of primary concern are the ESA listed endangered and threatened salmonids: Snake River fall-run Chinook salmon ESU (threatened); Snake River spring/summer-run Chinook salmon ESU (threatened); Lower Columbia River Chinook salmon ESU (threatened); Upper Columbia River spring-run Chinook salmon ESU (endangered); Columbia River chum salmon ESU (threatened); Snake River sockeye salmon ESU (endangered); Upper Columbia River steelhead ESU (endangered); Snake River Basin steelhead ESU (threatened); Lower Columbia River steelhead ESU (threatened); Middle Columbia River steelhead ESU (threatened). Listed fish can be impacted through a complex web of short and long term processes and over multiple time periods which makes evaluation of this a net effect difficult. WDFW is unaware of studies directly evaluating adverse ecological effects to listed salmon. In addition the program may have unknown impacts on eulachon populations in the basin.
- (3) Salmonid and non-salmonid fishes or other species that could positively impact the program. Multiple programs including fall Chinook, coho and steelhead programs are released from the North Toutle Hatchery and limited natural production of Chinook, coho, chum and steelhead occurs in this system along with non-salmonid fishes (sculpins, lampreys and sucker etc.).
- (4) Salmonid and non-salmonid fishes or species that could be positively impacted by the program. Coho smolts can be preyed upon release thru the entire migration corridor from the river sub-basin to the mainstem Columbia River and estuary. Northern pikeminnows and introduced spiny rays in the Columbia mainstem sloughs can prey on coho smolts as well as avian predators, including gulls, mergansers, cormorants, belted kingfishers, great blue herons and night herons. Mammals that benefit from migrating smolts and returning adults include: harbor seals, sea lions, river otters and orcas. Except for yearling coho and steelhead, these species may serve as prey items during the emigration through the basin. Hatchery fish provide an additional food source to natural predators that might otherwise consume listed fish and may overwhelm established predators providing a beneficial, protective effect to cooccurring wild fish. Hatchery releases can also behaviorally encourage mass emigration of multiple species through the watershed, reducing residency. Many watersheds in the Pacific Northwest appear to be nutrient-limited (Gregory et al. 1987; Kline et al. 1997) and salmonid carcasses can be an important source of marine derived nutrients (Levy 1997). Carcasses from returning adult salmonids have been found to elevate stream productivity through several pathways, including:
  - a) the releases of nutrients from decaying carcasses has been observed to stimulate primary productivity (Wipfli et al. 1998);
  - b) the decaying carcasses have been found to enrich the food base of aquatic invertebrates (Mathisen et al. 1988); and

c) Juvenile salmonids have been observed to feed directly on carcasses (Bilby et al. 1996).

#### 4 SECTION 4. WATER SOURCE

4.1 Provide a quantitative and narrative description of the water source (spring, well, surface), water quality profile, and natural limitations to production attributable to the water source.

**Table 4.1.1:** Water sources for the North Toutle Hatchery.

Water	Water Right		Available	Avg Water		
Source	Record/Cert No.	Permit No.	Water Flow	Temp. (F°) <sup>a</sup>	Usage	Limitations <sup>b</sup>
Green River (surface)	S2- *08904CWRIS/ 05609	08245	15 cfs	46 (32-70)	Incubation, rearing, acclimation, adult holding	Temps in lower river can reach in
Green River (surface) gravity intake	S2-24831 CWRIS		20 cfs		Rearing, acclimation	the 70s°F in the summer
Green River (surface) pump	S2-23796 CWRIS		4 cfs		Adult trapping pond	
Green River (surface)	S2-23797 CWRIS		9 cfs		Fishway	

Source: Phinney 2006, WDOE Water Resources Explorer 2014, WDFW hatchery data.

North Toutle Hatchery. A gravity intake on the Green River, located two miles from the confluence with the Toutle River, draws a maximum of 35cfs while a river pump adjacent to the adult trapping pond entrance supplies another 4 cfs. Water rights were obtained in 1949, 1975, 1976 and 1978.

The adult pond is 100% re-use water, and staff use aerators in order to boost oxygen levels in the adult pond and create some cover during August to October, if needed.

The raceway rearing ponds attain a maximum of 3,500 gpm total of gravity-fed water. On occasion, water temperatures in excess of 80°F have been observed during the summer months; temperatures in the 70s°F are routinely observed in the lower reaches of the Green River and hatchery rearing ponds (personal comm. Mark Johnson WDFW). The dissolved oxygen level of the incoming water decreases substantially during warm water periods and aerators are employed on the rearing ponds to maintain sufficient levels. Conversely, winter water temperatures may drop to freezing creating anchor and frazzle ice complicating water delivery.

#### **NPDES Permits:**

This facility operates under the "Upland Fin-Fish Hatching and Rearing" National Pollution Discharge Elimination System (NPDES) general permit which conducts effluent monitoring and reporting and operates within the limitations established in its permit administered by the Washington Department of Ecology (DOE).

Discharges from the cleaning treatment system are monitored as follows:

- *Total Suspended Solids (TSS)* 1 to 2 times per month on composite effluent, maximum effluent and influent samples.
- Settleable Solids (SS) 1 to 2 times per week on effluent and influent samples.
- In-hatchery Water Temperature daily maximum and minimum readings.

**Table 4.1.2:** Record of NPDES permit compliance.

Facility/	Reports Submitted Y/N		Last Inspection	Violations	Corrective	<b>Meets Compliance</b>	
Permit #	Monthly	Qtrly	Annual	Date	Last 5 yrs	Actions Y/N	Y/N
N Toutle WAG13-1010	Y	Y	Y	5/2/2006	0	N	Y

Source: Ann West, WDFW Hatcheries Headquarters Database 2013.

## 4.2 Indicate risk aversion measures that will be applied to minimize the likelihood for the take of listed natural fish as a result of hatchery water withdrawal, screening, or effluent discharge.

The North Toutle Hatchery intake structures were designed and constructed to specifications at the time the facility was built. The intake screens are in compliance with state and federal guidelines (NMFS 1995, 1996), but do not meet the current *Anadromous Salmonid Passage Facility Design* criteria (NMFS 2011). The pump intake sweep velocity, lack of fish bypass feature and 1/8 inch slotted screens are not in compliance (*Mitchell Act Intake and Fish Passage Study Report* 2003). The gravity intake screen slot width, approach and sweep velocities and an open top design into the intake (which allows high water to crest over this structure yearly), are not in compliance. WDFW has requested funding for scoping, design, and construction work of a new intake system. WDFW has completed a feasibility design report (2012) to address the intake compliance issues. These include:

- Upgrading the trash rack and intake screens to meet NMFS criteria, and to make them easier to clean and maintain;
- Raising or designing the walls to prevent overtopping during flood events; and
- Providing a fish bypass and outfall back to the Green River that is minimally attractive to upstream migrants;

#### 5 **SECTION 5. FACILITIES**

#### 5.1 Broodstock collection facilities (or methods).

Adult trapping begins in mid- to late-August. A temporary barrier "rack" is constructed across the Green River, approximately ¼-mile upstream of the confluence with the North Fork Toutle, to direct the fish to a trap located on the east side of the river. This rack is installed by August 20, and operates through late-November/early-December. Located on the right bank, the rack is flanked by the ladder section, which accesses the adult holding ponds. Picket sections (1.5 inch width) within the "rack" successfully blocks adults and all but the smallest jacks. The return is diverted into the holding ponds until the temporary weir is removed.

#### 5.2 Fish transportation equipment (description of pen, tank truck, or container used).

Adults are not transported for this program.

*SAFE program.* Green eggs are transferred in coolers via pick-up truck from North Toutle Hatchery to Grays River Hatchery in November (see Deep River Net Pens SAFE Coho HGMP). Sub-yearlings (25 fpp) were also transferred from North Toutle Hatchery to the Deep River Net Pens in November 2014; average transit time is 1.75 hours.

#### 5.3 Broodstock holding and spawning facilities.

**Table 5.3.1:** Adult holding/spawning facilities available.

Ponds (No.)	Pond Type	Volume (cu.ft)	Length (ft.)	Width (ft.)	Depth (ft.)	Available Flow (gpm)
2	Concrete holding ponds	10,800	60	40	4.5	2,000-3,000

The two concrete holding ponds 60 x 40 x 4.5 (ft.) are supplied with re-use water at 2,000-3,000 gpm. Aerators are used to increase oxygen levels and provide some cover to the adults. Spawning areas are located at the head end of the ponds with kill bins covered.

#### 5.4 Incubation facilities.

**Table 5.4.1:** Incubation vessels available.

Туре	Units (number)	Size	Flow (gpm)	Volume (cu.ft.)	Loading (eggs/unit)
Vertical Stack Tray Units	192 (8 tray stacks)	25" x 20 ¾" x 3.5"	3.5	0.22	7,000
Deep Troughs w/ incubation cells (8 cell/trough)	20	14.9° x 19" x 15"	5-12	29.5	225,000
Shallow Troughs	2	14.9' x 1' x 6.5"	5-9	8.0	20,000

### 5.5 Rearing facilities.

**Table 5.5.1:** Rearing facilities available.

Ponds (No.)	Pond Type	Volume (cu.ft)	Length (ft.)	Width (ft.)	Depth (ft.)	Flow (gpm)	Max. Flow Index	Max. Density Index
24	Concrete Raceways	3,638	80	20	2.3	150	1.61	0.20
1	Earthen Pond P-25	18,360	150	36	3.4	*	2.42	0.05
1	Earthen Pond P-26	98,832	426	58	4.0	*	2.42	0.05
1	Earthen Pond P-27	106,929	436	51	4.5	4,000	2.42	0.05
1	Earthen Pond P-28	36,693	151	54	4.5	1,500	2.42	0.05

<sup>\*</sup> Supplied from P-27

### 5.6 Acclimation/release facilities.

See HGMP section 5.5.

### 5.7 Describe operational difficulties or disasters that led to significant fish mortality.

- The eruption of Mount St. Helens in 1980 completely inundated the facility. Most of the hatchery infra-structure was renovated.
- A flood in February 1996 caused an unscheduled release of 110,000 coho fry.
- An epizootic of columnaris in 1996 resulted in a 60% mortality of the juvenile fingerling coho production.
- Low dissolved oxygen in the adult holding ponds in 2000, resulted in the loss of approximately 700 adult fall Chinook.

# 5.8 Indicate available back-up systems, and risk aversion measures that will be applied, that minimize the likelihood for the take of listed natural fish that may result from equipment failure, water loss, flooding, disease transmission, or other events that could lead to injury or mortality.

A prolonged loss of hatchery water supply would result in catastrophic loss of all rearing units, with incubation and the raceways being most vulnerable. Under a temporary cessation of the surface water supply, water can be re-directed from other supply sources as first pass or re-use to the units. Hatchery is staffed 24/7 and ready to react to system failure and WDFW has emergency procedures and plans in place. All systems are alarmed to alert us of failure.

IHOT fish health guidelines are followed. WDFW fish health specialists conduct inspections monthly and problems are managed promptly to limit mortality and reduce possible disease transmission. In the event of possible virus outbreak, WDFW facilities follow very strict disinfection procedures and comprehensive lab analysis of all egg-takes for culling, if needed.

### 6 SECTION 6. BROODSTOCK ORIGIN AND IDENTITY

Describe the origin and identity of broodstock used in the program, its ESA-listing status, annual collection goals, and relationship to wild fish of the same species/population.

#### 6.1 Source.

Adults returning to North Toutle Hatchery. Natural-origin fish (adipose intact) incorporation into the broodstock will be maximized to allow for integration of the on-station program.

### **6.2** Supporting information.

### **6.2.1 <u>History.</u>**

Coho are native to the Toutle River, were historically abundant and present throughout the watershed in all accessible tributaries. A major portion of the spawning area was destroyed by the eruption of Mt. St. Helens in 1980 (WDF et. al. 1993). Toutle River coho were generally an early-returning stock (Type-S), with most fish returning from August through October. Late-runs are also present. Adult coho are trapped and hauled above the sediment-retention dam on the North Fork Toutle River (WDW 1990). Hatchery fingerlings were seeded in the watershed beginning with the 1984 brood. Rearing ponds were developed near the original hatchery site after the eruption, and began operation in 1985 (LCFRB Plan 2010).

This broodstock was largely founded from native coho salmon in the Toutle River. There was a substantial wild coho salmon run into that basin when the hatchery opened. The hatchery fish were not marked, so levels of natural-origin fish in the broodstock are unknown. After the eruption of Mt. St. Helens and the destruction of the hatchery in 1980, adults returning to the North Forth Toutle River were captured and transported to Grays River Hatchery for rearing and release. Selective marking and subsequent spawning allowed this stock to be kept separate. The North Toutle Hatchery was renovated and began collecting broodstock again in 1990 (LCFRB Plan 2010).

### 6.2.2 Annual size.

*Integrated program.* Around 70 adult pairs, not including jacks, are needed to achieve the established egg-take goal of 200,000 (FBD 2014) for the on-station program. This is based on an average fecundity of around 3,000 smolts/female, and a pre-spawning mortality of 10%.

*Segregated program.* A total of around 335 hatchery-origin adult pairs are needed to achieve the egg-take goal of 910,000 collected for the Selected Area Fisheries Enhancement (SAFE) releases from Deep River Net Pens (see Deep River Net Pens SAFE Coho HGMP).

### 6.2.3 Past and proposed level of natural fish in broodstock.

All Type S coho at North Toutle Hatchery have been mass-marked since brood year 1995, release year 1997. The production has operated as a segregated type program, with no natural fish incorporated into the broodstock since 1998. WDFW began integrating the maximum number of available natural spawners into the broodstock program in 2006. The pNOB for the program over the last three years is 1.0. See **Table 7.4.2.1**.

### 6.2.4 Genetic or ecological differences.

All adults recruited for broodstock have been collected at the facility since 1998. There are no known genotypic, phenotypic, or behavioral differences between either the hatchery or natural stocks in the sub-basin. This broodstock was founded from Toutle River coho, and has received few non-ESU or out-of-basin fish since 1998. Although there may have been historical releases of non-ESU fish into the Toutle sub-basin, they were not used to establish the broodstock.

### 6.2.5 Reasons for choosing.

Locally available stock.

The Type S coho in the Toutle River, best represents the stock timing that provides harvest opportunities for fisheries in the sub-basin, the lower Columbia mainstem/tributaries, Washington/Oregon Coast. The early stocks are the strength of the Buoy 10 coho fishery at the mouth of the Columbia River. They also return to the tributaries when the weather is warmer and stream flows are moderate, providing excellent sport fishing opportunities.

## 6.3 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic or ecological effects to listed natural fish that may occur as a result of broodstock selection practices.

- Natural spawners will be integrated into the broodstock to represent the natural Type S coho run throughout the season.
- Hatchery program fish are mass-marked.
- There are no known genotypic, phenotypic, or behavioral differences between either the hatchery stock or natural stock in the sub-basin.
- Holding pond procedures follow IHOT guidelines.
- Other listed fish encountered during the broodstock collection process passed into the upper watershed, with minimal handling and holding time.
- Any observed mortalities will be reported in the WDFW Hatcheries Headquarters Database.

### 7 SECTION 7. BROODSTOCK COLLECTION

### 7.1 Life-history stage to be collected (adults, eggs, or juveniles).

Adults returning to the North Toutle Hatchery.

### 7.2 Collection or sampling design.

"Type S" coho are collected each year from the run at large reaching North Toutle Hatchery weir trap during September and early Decmeber. Broodstock are collected throughout the entire run to maintain run timing for the population. Capture efficiency is 100% for fish volunteering to the trap. The river is blocked with a temporary picket barrier (in place from late-August through December) upstream and adjacent to the ladder exiting the trap, which prevents migration upstream of the capture point.

### 7.3 Identity.

All Type-S coho produced from this facility have been released mass-marked with an adipose finclip (AD) since the 1995 brood year. A portion of the on-station releases (~20%) are also released AD+coded-wire tagged (CWT). Only AD-marked fish were included in the broodstock from 1998 through 2005; unmarked (presumed wild) fish have been integrated into the broodstock since 2006.

### 7.4 Proposed number to be collected:

### 7.4.1 **Program goal (assuming 1:1 sex ratio for adults):**

See HGMP section 6.2.2.

### 7.4.2 <u>Broodstock collection levels for the last twelve years, or for most recent years available:</u>

**Table 7.4.2.1:** Composition of marked and unmarked adults and jacks in the hatchery broodstock.

Dunad Wasa	Marked			Unmarked		
Brood Year	Females	Males	Jacks	Females	Males	Jacks
2004	288	288	1			
2005	367	369	6			
2006	301	282	2	7	11	0
2007	303	266	1	34	63	0
2008	47	45	1	9	11	0
2009	39	18	2	28	48	1
2010	31	20	1	21	31	0
2011				66	63	3
2012				71	68	3
2013				58	55	3

Source: WDFW Hatcheries Headquarters Database 2014.

Note: Egg-take was reduced in 2008 in response to CS&F plan.

See also Deep River Net Pen (SAFE) Type-S Coho HGMP for segregated program broodstock collection levels.

### 7.5 Disposition of hatchery-origin fish collected in surplus of broodstock needs.

In addition to the adults needed to meet broodstock goals, adults may be released upstream of the rack in the Green River but most stay within pHOS guidelines. Additional surplus may be donated to local food bank organizations or used for nutrient enhancement.

### 7.6 Fish transportation and holding methods.

Adults are not transported for this program.

### 7.7 Describe fish health maintenance and sanitation procedures applied.

The adult holding area is separated from all other hatchery operations. Disinfection procedures that prevent pathogen transmission between stocks of fish are implemented during spawning. Spawning implements are rinsed with an iodophor solution, and spawning area and implements are disinfected with iodophor solution at the end the spawning day.

### 7.8 Disposition of carcasses.

Spawned carcasses not used for system nutrient enhancement will be disposed of in a landfill.

## 7.9 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic or ecological effects to listed natural fish resulting from the broodstock collection program.

Proper trap operation and fish handling techniques are followed. Out-of-basin transfers are limited. Broodstock are collected throughout the return period. Broodstock collection procedures

quickly identify non-target fish encountered; fish not used in the program are released immediately.

### 8 SECTION 8. MATING

Describe fish mating procedures that will be used, including those applied to meet performance indicators identified previously.

#### 8.1 Selection method.

Representative portions of the run are taken weekly, and spawning occurs over a period of up to six weeks, from early-October to early November. Spawning activity peaks between October 20 and November 1.

### **8.2** Males.

A ratio of 1:1 males to females is used. Jack coho salmon (2-year old) are incorporated into the broodstock as males at a minimum of 5% of the total spawning population.

### 8.3 Fertilization.

Agency spawning guidelines are closely followed (Seidel 1983). Fertilization occurs at a 1:1 ratio (females/males). Milt is mixed with green eggs with the ovarian fluid. Water hardening procedures with iodophor are followed after twenty minutes. Iodophor solution is used as rinse that is applied to hands and spawning implements per spawning. Iodophor foot baths are located at entrance to incubation room. Generally, sixty ovarian fluid and kidney/spleen samples are collected from female spawners to test for the presence of viral pathogens.

### 8.4 Cryopreserved gametes.

Cryopreserved gametes are not used.

## 8.5 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic or ecological effects to listed natural fish resulting from the mating scheme.

- Mating cohorts are randomly selected.
- Protocols for population size, fish health disinfection and genetic guidelines followed.
- Spawn all collected mature broodstock if possible without regard to age, size, color or other physical characteristics. If not spawning all collected mature adults over the season, apply the same rationale to individual spawn days.
- Randomize mating and avoid selectivity beyond ripeness on a given spawn day.
- Use one male to one female as much as possible in order to ensure an equal genetic contribution.
- Do not mix milt from multiple males and add to eggs (pooling prior to mixing) in order to eliminate disproportionate genetic male contributions.
- Do not re-use males except as part of specific spawning protocols. A given male should be used as the first mate for only one female total.

9 <u>SECTION 9. INCUBATION AND REARING</u>-Specify any management *goals* (e.g. "egg to smolt survival") that the hatchery is currently operating under for the hatchery stock in the appropriate sections below. Provide data on the success of meeting the desired hatchery goals.

### 9.1 Incubation:

### 9.1.1 Number of eggs taken and survival rates to eye-up and/or ponding.

**Table 9.1.1.1:** Survival rates (%) from egg-take to ponding.

Brood Year	Faa Toko	% Egg	g Survival
brood rear	Egg Take	Green-to-Eyed	Eyed Egg-to-Ponding
2004	1,033,000	91.3	97.2
2005	1,039,403	88.1	97.0
2006	1,000,000	91.4	99.0
2007	996,542	88.9	100.0
2008	218,252	83.4	100.0
2009	216,756	90.4	99.8
2010	203,447	87.2	99.0
2011	228,627	94.1	98.0
2012	216,534	95.1	99.0
2013	218,328	94.8	99.0

Source: WDFW hatchery records 2014.

See Deep River Net Pen SAFE Coho HGMP for segregated program egg-take.

### 9.1.2 Cause for, and disposition of surplus egg takes.

Egg-takes can be heavily-loaded to the first part of the run if escapement appears to be unlikely at the beginning of the season due to low water, environmental conditions during September and early October. To preserve later run timed takes, early eggs can be disposed of after consultation with region staff at a landfill.

In the event that egg survival is higher than expected, WDFW Regional Managers will be contacted for instructions for disposition of the surplus in accordance with Regional policy and guidelines set forth in management plans/agreements and ESA permits.

### 9.1.3 Loading densities applied during incubation.

Eggs are placed in deep troughs to the eye stage then moved to stack incubators for hatching. Removal of dead eggs, accurate enumeration and loadings are adjusted during this time (see HGMP section 5.4 for loading criteria). WDFW follows *Integrated Hatchery Operations Team* (IHOT) species-specific incubation recommendations for water quality, flows, temperature, substrate, and incubator capacities.

### 9.1.4 <u>Incubation conditions.</u>

IHOT species-specific incubation recommendations are followed for water quality, flows, temperature, substrate and incubator capacities. Harmful silt and sediment is cleaned from incubation systems regularly while eggs are monitored to determine fertilization and mortality. Incubation water temperature is monitored by thermograph and recorded and temperature units (TU) are tracked for embryonic development. Dissolved oxygen content is monitored and ranged from 8-12 parts per million (ppm). When using artificial substrate, Vexar® or bio-rings, egg densities within incubation units are reduced by 10%.

### **9.1.5 Ponding.**

Fry are typically ponded mid-February through early April, when the yolk slit is closed to approximately 1-mm wide (approximately 1320 TUs) or KD factor (95% yolk absorption). Fry are transferred by irrigation piping from the incubation room to the appropriate raceway.

### 9.1.6 Fish health maintenance and monitoring.

Staff conducts daily inspection, visual monitoring and sampling from eye, fry fingerling and subyearling stages. As soon as potential problems are seen, these concerns are immediately communicated to the WDFW fish health specialist. In addition, fish health specialists conduct inspections monthly. Potential problems are managed promptly to limit mortality and reduce possible disease transmission. Disease treatment varies with the pathogen encountered, but is generally antibiotic in nature for bacterial infections and bath or drip treatments with chemotheraputants for external infections

## 9.1.7 <u>Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish during incubation.</u>

- IHOT and WDFW fish health guidelines followed.
- Multiple units are used in incubators.
- Splash curtains are not used at this facility, but can be installed to isolate incubators.
- Temperature, dissolved oxygen, and flow are monitored.
- Dead eggs are discarded in a manner that prevents disease transmission.

### 9.2 Rearing:

## 9.2.1 Provide survival rate data (average program performance) by hatchery life stage (fry to fingerling; fingerling to smolt) for the most recent twelve years (1988-99), or for years dependable data are available.

**Table 9.2.1.1:** Survival rates (%) from ponding to release.

Brood	% Survival			
Year	Fry-to-Sub-yearling	Sub-yearling-to-Smolt		
2004	79.6	94.8		
2005	85.1	100.0		
2006	91.7	100.0		
2007	92.5	100.0		
2008	80.9	100.0		
2009	89.3	94.5		
2010	93.2	94.9		
2011	81.3	95.3		
2012	88.9	91.0		
2013	n/a	n/a		

Source: Hatcheries Headquarters Database 2014, hatchery records.

NA – Not available

<sup>&</sup>lt;sup>a</sup> A total of 356,388 sub-yearlings were transferred to the Deep River Net Pen SAFE program in November 2008.

### 9.2.2 <u>Density and loading criteria (goals and actual levels).</u>

Loading and density levels at WDFW hatcheries conform to standards and guidelines set forth in *Fish Hatchery Management* (Piper et. al. 1982), the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006). IHOT standards are followed for water quality, alarm systems, predator control measures to provide the necessary security for the cultured stock, loading and density.

Densities are kept at or below 3.3 lbs/gpm and 0.5 lbs/cu ft. before the last loading reduction in the fall of the year. Trough maximum loading is 40 lbs at 12 gpm (3.33 lbs/gpm). Tank and raceway maximum loading for early rearing is 132 lbs for the tanks at 40 gpm (3.3 lbs/gpm) and 800 lbs per raceway at 300 gpm.(2.66 lbs/gpm). The final loading per raceway is approximately 3,200 lbs at 300 gpm (10.6 lbs/gpm), or 10,000 lbs at 1,500 gpm in the rearing ponds (6.67 lbs/gpm).

### 9.2.3 Fish rearing conditions

**Table 9.2.3.1:** Monthly average surface water temperature (°F) at North Toutle Hatchery.

Month	Average Water Temperature (°F)
January	39
February	40
March	41
April	44
May	47
June	52
July	62
August	62
September	57
October	48
November	43
December	40

Source: WDFW Hatchery Records 2014.

IHOT standards are followed for water quality, alarm systems, predator control measures (netting), loading and density.

Fish are reared on river water. Temperature, dissolved oxygen (DO) and pond turn-over rate are monitored and recorded daily during fish rearing; water temperatures during the rearing cycle range from 32°F to 80°F (**Table 9.2.3.1**). Ponds are vacuum-cleaned regularly, generally weekly, to remove settleable solids, unused feed and feces to ensure proper cleanliness. Predator netting over the rearing ponds minimize predation. All ponds are pressure washed between broods.

Fish are mass-marked in June when they are about 120 fpp.

## 9.2.4 <u>Indicate biweekly or monthly fish growth information (average program performance)</u>, including length, weight, and condition factor data collected during rearing, if available.

**Table 9.2.4.1:** Monthly fish growth information by length (mm), weight (fpp), condition factor and growth rate, collected during rearing.

Rearing Period	Length (mm)	Weight (fpp)	Growth Rate
March	40.3	847	Na

April	50.2	432	0.490
May	64.7	196	0.546
June	75.9	127	0.352
July	77.1	120	0.055
August	83.8	90	0.250
September	93.1	68	0.244
October	110.0	40	0.412
November	114.9	36	0.100
December	123.1	29	0.194
January	125.0	28	0.035
February	133.7	23	0.179
March	124.0	20	Na
April	136.0	15	Na

Source: WDFW Hatchery Records.

### 9.2.5 <u>Indicate monthly fish growth rate and energy reserve data (average program performance)</u>, if available.

See HGMP section 9.2.4. No energy reserve data available.

## 9.2.6 Indicate food type used, daily application schedule, feeding rate range (e.g. % B.W./day and lbs/gpm inflow), and estimates of total food conversion efficiency during rearing (average program performance).

Fish are given variety of diet formulations including starter, crumbles and pellets; the food brand used may vary, depending on cost and vendor contacts. Feeding frequencies varies depending on the fish size and water temperature, and usually begin at 7-8 feedings/7 days a week, and end at 1-3 feedings/7 days a week. Fall Chinook are typically fed around 1.5-1.8% B.W./day, depending upon water temperature and weather conditions. Feed rate is applied in accordance with program goals not to exceed 0.1-0.15 pounds feed per gallon in-flow, depending on fish size. Average season conversion rates generally are no greater than 1.3:1.0.

### 9.2.7 Fish health monitoring, disease treatment, and sanitation procedures.

Monitoring. Policy guidance includes: Fish Health Policy in the Columbia Basin. Details hatchery practices and operations designed to stop the introduction and/or spread of any diseases within the Columbia Basin. Also, Policies and Procedures for Columbia Basin Anadromous Salmonid Hatcheries (Fish Health Policy Chapter 5, IHOT 1995). A fish health specialist inspects fish monthly and checks both healthy and presence of symptomatic fish. Based on pathological or visual signs by the crew, age of fish and the history of the facility, the pathologist determines the appropriate tests. External signs such as lesions, discolorations, and fungal growths will lead to internal examinations of skin, gills and organs. Blood is checked for signs of anemia or other pathogens. Additional tests for virus or parasites are done if warranted.

Disease Treatment. As needed, appropriate therapeutic treatment will be prescribed to control and prevent further outbreaks. Formalin treatments are used against Costia during April and June. Outbreaks of columnaris, furunculosis and Cold Water Disease (CWD) can occur and are treated with potassium permanganate baths, Terramycin®, and prophylactic immune responses (proactive immunostimulant). Problems usually subside with cooler temperatures in October. Mortality is collected and disposed of at a landfill. Fish health and or treatment reports are kept on file. Mortality is collected and disposed of at a landfill. Fish health and or treatment reports are kept on file.

Sanitation. All eggs brought to the facility are surface-disinfected with iodophor (as per disease policy). All equipment (nets, tanks, boots, etc.) is disinfected with iodophor between different fish/egg lots. Different fish/egg lots are physically isolated from each other by separate ponds or incubation units. The intent of these activities is to prevent the horizontal spread of pathogens by splashing water. Tank trucks are disinfected between the hauling of adult and juvenile fish. Foot baths containing disinfectant are strategically located on the hatchery grounds to prevent spread of pathogens.

### 9.2.8 Smolt development indices (e.g. gill ATPase activity), if applicable.

ATPase gill activity is not measured. Fish size at release time is critical to the readiness for migration. The migratory state of the release population is determined by fish behavior. Aggressive screen and intake crowding, swarming against sloped pond sides, a leaner (0.80 – 0.90) condition factor (K), a silvery physical appearance and loose scales during feeding events are signs of smolt development. Surface water from the Green River is used for fish rearing, and provides a natural water temperature profile.

### **9.2.9** <u>Indicate the use of "natural" rearing methods as applied in the program.</u> Not applicable.

## 9.2.10 <u>Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish under propagation.</u>

See HGMP sections 5.8, 6.3, 7.9 and 9.1.7.

### 10 SECTION 10. RELEASE

Describe fish release levels, and release practices applied through the hatchery program.

### 10.1 Proposed fish release levels.

**Table 10.1.1**: Proposed release levels (maximum number).

Age Class	Maximum Number	Size (fpp)	Release Date	Location
Yearling	150,000	15.0	May	Cowlitz

Source: WDFW Future Brood Document 2014.

Note: 15 fpp = 146 mm fork length (fl).

This does not include the Deep River Net Pen coho releases (see Deep River Net Pen SAFE Coho HGMP).

### 10.2 Specific location(s) of proposed release(s).

**Stream, river, or watercourse:** Green River (WRIA 26.0323)

**Release point:** RKm 0.81

Tributary to NF Toutle River (WRIA 26.0314) at R.M. 11.5

**Major watershed:** Cowlitz

**Basin or Region:** Lower Columbia

### 10.3 Actual numbers and sizes of fish released by age class through the program.

**Table 10.3.1:** Number of fish released, size, CVs and release date, by age and year.

Dalaasa Vaan	Yearlings				
Release Year	Number	Avg Size (fpp)	CV	Date	
2002	986,490	15.4	4.92	April 26-May 15	
2003	808,100	14.8	4.63	May 1-15	
2004	769,926	14.7	5.80	May 5-12	
2005	597,750	16.4	6.70	April 28-May 6	
2006	691,881	13.7	5.60	May 12-24	
2007	755,112	15.3	5.10	May 1-24	
2008	829,666	14.9	5.95	May 15-30	
2009	489,641	15.0	5.64	May 15-30	
2010	147,232	14.2	5.11	May 3-10	
2011	164,985	14.9	5.62	May 2-9	
2012	155,310	14.9	5.41	May 1-8	
2013	163,289	14.7	5.50	May 2-9	

Source: WDFW Hatcheries Headquarters Database 2014.

Note: 13.7 fpp = 151 mm fork length (fl); 14.5 fpp = 148 mm fl; 15 fpp = 146 mm fl; 16 fpp = 143 mm fl

### 10.4 Actual dates of release and description of release protocols.

Volitional releases generally start on May 1, and continue through May 15. Ponds screens are removed first, and then stop-logs are removed over the next few days to slowly lower the pond. Most of the fish vacate the pond after the first week. Most of the stop-logs are removed by the second week. See **Table 10.3.1** for actual release dates.

### 10.5 Fish transportation procedures, if applicable.

This is an-station release; juvenile fish are not transported.

*SAFE program.* Around 225,000 sub-yearlings are shipped to the Deep River net pens program in November; (see Deep River Net Pen SAFE Coho HGMP).

### 10.6 Acclimation procedures (methods applied and length of time).

Fish are reared and acclimated on river water during their entire time at North Toutle Hatchery. Yearlings are released directly from the ponds into the Green River.

### 10.7 Marks applied, and proportions of the total hatchery population marked, to identify hatchery adults.

**Table 10.7.1:** Marks applied, by brood year, age class and mark-type, North Toutle Hatchery onstation releases.

Brood Year	Yearlings	Mark Type
2014	45,000	AD+CWT
2014	117,000	AD-only

Fish are 100% mass-marked with either adipose fin-clips + coded-wire tag (AD+CWT) or AD-only, to differentiate them from the natural population.

Snouts collected from the adipose fin-clipped adults are dissected, recovered and read at the WDFW CWT Lab in Olympia. Scale samples are read at WDFW Headquarters Olympia to verify hatchery- or natural-origin.

### 10.8 Disposition plans for fish identified at the time of release as surplus to programmed or approved levels.

The program guidelines for annual broodstock/egg-take collection are managed to prevent any surpluses, and maintained within the  $\pm 5\%$  guideline. In the event of surplus >10%, WDFW Regional Managers will in accordance with regional policy and guidelines set forth in management plans/agreements and ESA permits, and after consultation with NMFS, instruct hatchery staff for disposition of the surplus.

### 10.9 Fish health certification procedures applied pre-release.

All fish are examined for the presence of "reportable pathogens" as defined in the Pacific Northwest Fish Health Protection Committee (PNFHPC) disease control guidelines, within three weeks prior to release.

Fish transfers into the sub-basin are inspected and accompanied by notifications as described in IHOT and PNFHPC guidelines.

Prior to release, the population health and condition is established by the Area Fish Health Specialist. This is commonly done 1-3 weeks pre-release and up to 6 weeks on systems with pathogen-free water and little or no history of disease. Prior to this examination, whenever abnormal behavior or mortality is observed, staff also contacts the Area Fish Health Specialist. The fish specialist examines affected fish, and recommends the appropriate treatment. Reporting and control of selected fish pathogens are done in accordance with the *Salmonid Disease Control Policy of the Fisheries Co-Managers of Washington State* (WDFW and WWTIT 1998, updated 2006) and IHOT guidelines.

### 10.10 Emergency release procedures in response to flooding or water system failure.

Hatchery staff will make every attempt to keep the fish alive and healthy throughout the entire rearing-release cycle; all appropriate resource managers from the Complex level to the Federal level will be informed of the actions taken. Based in authorized instructions, screens and stoplogs could be pulled for direct/forced fish release into the Green River.

### 10.11 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish resulting from fish releases.

- The production and release of only smolts through fish culture and volitional release practices fosters rapid seaward migration with minimal delay in the rivers, limiting interactions with naturally produced juveniles.
- Coho smolts are released in May to allow listed Chinook to grow to a size that will reduce predation opportunities, and still be in advance of winter and summer steelhead fry emergence in Columbia River tributaries.
- Returning hatchery fish are under heavy selective harvest, and may be differentiated from natural-origin fish by the adipose fin-clip or CWT.
- WDFW proposes to continue monitoring, research and reporting of hatchery smolt migration performance behavior, and intra and interspecific interactions with wild fish to access, and adjust if necessary, hatchery production and release strategies to minimize effects on wild fish.
- WDFW fish health and operational concerns for North Toutle Hatchery programs are communicated to WDFW Region 5 staff for any risk management or needed treatment. See also HGMP section 9.7.

### 11 <u>SECTION 11. MONITORING AND EVALUATION OF</u> PERFORMANCE INDICATORS

### 11.1 Monitoring and evaluation of "Performance Indicators" presented in Section 1.10.

### 11.1.1 <u>Describe plans and methods proposed to collect data necessary to respond to each "Performance Indicator" identified for the program.</u>

Performance indicators for harvest will be accomplished by continuing mass marking (ad clip). CWT recoveries will help determine stray rate contributions on spawning grounds by watersheds close in proximity to this program's release vicinity. See HGMP section 1.10 Monitoring and Evaluation for additional plans and methods to collect data necessary.

Additional research, monitoring and evaluation in the Lower Columbia. WDFW is currently conducting the following Mitchell Act-funded research, monitoring and evaluation projects:

**Table 11.1.1.1:** Current WDFW Mitchell Act-funded research, monitoring and evaluation projects.

Project Project	Description
LCR Monitoring	WDFW has implemented an expanded monitoring program for Chinook, coho, chum and steelhead populations in the Lower Columbia River (LCR) region of Southwest Washington (WDFW's Region 5) and fishery monitoring in the lower mainstem of the Columbia River. The focus of this expanded monitoring is to 1) gather data on Viable Salmonid Population (VSP) parameters – spawner abundance, including proportion of hatchery-origin spawners (pHOS), spatial distribution, diversity, and productivity, 2) to increase the coded wire tag (CWT) recovery rate from spawning grounds to meet regional standards, and 3) to evaluate the use of PIT tags to develop harvest rates for salmon and steelhead populations. Additionally, key watersheds are monitored for juvenile salmonid out-migrant abundance. Coupled with adult abundance information, these data sets allow for evaluation of freshwater productivity and development of biological reference points, such as seeding capacity. Monitoring protocols and analysis methods utilized are intended to produce unbiased estimates with measurements of precision in an effort to meet NOAA monitoring guidelines (Crawford and Rumsey 2011).

## 11.1.2 <u>Indicate whether funding, staffing, and other support logistics are available or committed to allow implementation of the monitoring and evaluation program.</u>

Except for a risk involving genetic introgression, all other aspects of the M&E outlined in HGMP section 1.10 are currently funded (see also HGMP section 11.1.1).

## 11.2 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse genetic and ecological effects to listed fish resulting from monitoring and evaluation activities.

Monitoring, evaluation and research follow scientific protocols with adaptive management process if needed. WDFW will take risk aversion measures to eliminate or reduce ecological effects, injury, or mortality as a result of monitoring activities See HGMP section 1.10 Monitoring and Evaluation for additional plans and methods to collect data necessary, In addition, we will adaptively manage all aspects of the program to continue to minimize associated risks using the more recent available scientific research.

### 12 SECTION 12. RESEARCH

12.1 Objective or purpose.

No research is directly associated with the program.

12.2 Cooperating and funding agencies.

Any research is conducted by WDFW and funded through Tacoma Power.

12.3 Principle investigator or project supervisor and staff.

Cowlitz Hatchery Biologist

12.4 Status of stock, particularly the group affected by project, if different than the stock(s) described in Section 2.

Not applicable.

12.5 Techniques: include capture methods, drugs, samples collected, tags applied.

Not applicable.

12.6 Dates or time period in which research activity occurs.

Not applicable.

12.7 Care and maintenance of live fish or eggs, holding duration, transport methods.

Not applicable.

12.8 Expected type and effects of take and potential for injury or mortality.

Not applicable.

12.9 Level of take of listed fish: number or range of fish handled, injured, or killed by sex, age, or size, if not already indicated in Section 2 and the attached "take table" (Table 1).

Not applicable.

12.10 Alternative methods to achieve project objectives.

Not applicable.

12.11 List species similar or related to the threatened species; provide number and causes of mortality related to this research project.

Not applicable.

12.12 Indicate risk aversion measures that will be applied to minimize the likelihood for adverse ecological effects, injury, or mortality to listed fish as a result of the proposed research activities.

Not applicable.

### 13 <u>SECTION 13. ATTACHMENTS AND CITATIONS</u>

Beamesderfer, R., L. Berg, M. Chilcote, J. Firman, E. Gilbert, K. Goodson, D. Jepsen, T. Jones, S. Knapp, C. Knutsen, K. Kostow, B. McIntosh, J. Nicholas, J. Rodgers, T. Stahl, and B. Taylor. 2010. Lower Columbia River conservation and recovery plan for Oregon populations of salmon and steelhead. Oregon Department of Fish and Wildlife. 423 pp. Salem, Oregon. Available from: <a href="http://www.dfw.state.or.us/fish/CRP/docs/lower-columbia/OR\_LCR\_Plan%20-%20Aug\_6\_2010\_Final.pdf">http://www.dfw.state.or.us/fish/CRP/docs/lower-columbia/OR\_LCR\_Plan%20-%20Aug\_6\_2010\_Final.pdf</a>.

Bilby R.E., B.R. Fransen, and P.A. Bisson. 1996. Incorporation of nitrogen and carbon from spawning coho salmon into the trophic system of small streams: evidence from stable isotopes. Canadian Journal of Fisheries and Aquatic Sciences 53:164–173.

Crawford, B.A. and S. Rumsey. 2011. Guidance for Monitoring Recovery of Pacific Northwest Salmon & Steelhead listed under the Federal Endangered Species Act (Idaho, Oregon, and Washington. NMFS NW Region. January 2011.

Dornbusch, P. and A. Sihler. 2013. ESA recovery plan for Lower Columbia River coho salmon, Lower Columbia River Chinook salmon, Columbia River chum salmon, and Lower Columbia River steelhead. National Marine Fisheries Service. Northwest Region, Portland, Oregon. 503 pp.

Gustafson, R.G., M.J. Ford, D. Teel, and J.S. Drake. 2010. Status review of eulachon (*Thaleichthys pacificus*) in Washington, Oregon, and California. U.S. Dept. Commer., NOAA Tech. Memo. NMFS-NWFSC-105, 360 p.

Ford M.J. (ed.). 2011. Status review update for Pacific salmon and steelhead listed under the Endangered Species Act: Pacific Northwest. U.S. Dept. Commer., NOAA Tech. Memo. NMFS-NWFSC-113, 281 p.

Good, T.P., R.S. Waples, and P. Adams, (editors). 2005. Updated status of federally listed ESUs of West Coast salmon and steelhead. U.S. Department Commerce. NOAA Tech. Memo. NMFS-NWFSC-66.

HSRG (Hatchery Scientific Review Group). 2004. Hatchery reform; principles and recommendations of the Hatchery Scientific Review Group. Long Live the Kings. Seattle, Washington. Available from: <a href="http://hatcheryreform.us/hrp\_downloads/reports/hsrg\_princ\_recs\_report\_full\_apr04.pdf">http://hatcheryreform.us/hrp\_downloads/reports/hsrg\_princ\_recs\_report\_full\_apr04.pdf</a>

HSRG (Hatchery Scientific Review Group). 2009. Report to Congress on Columbia River Basin Hatchery Reform. Hatchery Scientific Review Group. Long Live the Kings. Seattle, Washington.

http://hatcheryreform.us/hrp\_downloads/reports/columbia\_river/report\_to\_congress/hsrg\_rep\_ort\_12.pdf

HSRG (Hatchery Scientific Review Group). 2009. Hatchery Scientific Review Group review and recommendations: Columbia Estuary – Deep River Net Pen coho population and related Hatchery Programs. Long Live the Kings. Seattle, Washington. Available from: <a href="http://hatcheryreform.us/hrp/reports/system/welcome\_show.action">http://hatcheryreform.us/hrp/reports/system/welcome\_show.action</a>.

IHOT (Integrated Hatchery Operations Team). 1995. Operation plans for anadromous fish Production facilities in the Columbia River basin. Volume III - Washington. Annual Report 1995. Bonneville Power Administration, Portland, OR. Project Number 92-043. 536 pp.

LCFRB (Lower Columbia Fish Recovery Board). 2010. Lower Columbia Salmon Recovery and Fish & Wildlife Subbasin Plan. June 6, 2010.

http://www.lcfrb.gen.wa.us/Recovery%20Plans/March%202010%20review%20draft%20RP/RP%20Frontpage.htm.

Mathisen, O.A., P.L. Parker, J.J. Goering, T.C. Kline, P.H. Poe, and R.S. Scalan. 1988. Recycling of marine elements transported into freshwater systems by anadromous salmon. Verh. Int. Ver. Limnol. 23: 2249-2258.

McElhany, P., M.H. Ruckelhaus, M.J. Ford, T.C. Wainwright, and E.P. Bjorkstedt. 2000. Viable salmonid populations and the recovery of evolutionarily significant units. U.S. Dept. Commerce, NOAA Tech. Memo. NMFS-NWFSC-42.

McElhany, P., C. Busack, M. Chilcote, S. Kolmes, B. McIntosh, J. Myers, D. Rawding, A. Steel, C. Steward, D. Ward, T. Whiesel, C. Willis. 2006. Revised viability criteria for salmon and steelhead populations in the Willamette and lower Columbia basins, review draft. Willamette/Lower Columbia Technical Recovery Team (WLC-TRT) and Oregon Department of Fish and Wildlife (ODFW). Portland, Oregon.

McElhany, P., M. Chilcote, J. Myers, R. Beamesderfer. 2007. Viability status of Oregon salmon and steelhead populations in the Willamette and lower Columbia basins, review draft. NMFS-NWFSC. Seattle, Washington.

McElhany, P., T.Bachman, C. Busack, S. Heppell, S. Kolmes, A. Maule, J. Myers, D. Rawding, D. Shively, A. Steel, C. Steward, and T. Whitesel. 2003. Interim report on viability criteria for Willamette and Lower Columbia Basin Pacific salmonids. Unpublished report. NOAA Fisheries.

Myers, J., C. Busack, D. Rawding, A. Marshall, D. Teel, D.M. Van Doornik, and M.T. Maher. 2006. Historical population structure of Willamette and Lower Columbia River Basin Pacific salmonids. United States Department of Commerce. NOAA Technical Memorandum NMFS-NWFSC-73. Seattle, Washington.

NMFS (National Marine Fisheries Service). 1995. Juvenile fish screen criteria for pump intakes. Available from: <a href="http://www.nwr.noaa.gov/1hydrop/nmfscrit1.htm">http://www.nwr.noaa.gov/1hydrop/nmfscrit1.htm</a>.

NMFS (National Marine Fisheries Service). 1996. Juvenile fish screen criteria for pump intakes. Available from: http://www.nwr.noaa.gov/1hydrop/pumpcrit1.htm.

NMFS (National Marine Fisheries Service). 1999. Endangered and threatened species: Threatened status for three Chinook salmon Evolutionarily Significant Units in Washington and Oregon, and Endangered status for one Chinook salmon ESU in Washington; final rule. Partial 6-month extension on final listing determinations for four Evolutionarily Significant Units of West Coast Chinook salmon; proposed rule. Federal Register 64:14308-14328.

NMFS (National Marine Fisheries Service). 2000a. A risk assessment procedure for evaluating harvest mortality of Pacific salmonids. National Marine Fisheries Service, Sustainable Fisheries Division, Northwest Region. May 30. 33pp.

NMFS (National Marine Fisheries Service). 2005. Endangered and threatened species: final listing determinations for 16 ESUs of west coast salmon, and final 4(d) protective regulations for threatened salmonid ESUs. Federal Register 70FR37160.

NMFS (National Marine Fisheries Service). 2006. Endangered and Threatened Species: Final Listing Determinations for 10 Distinct Population Segments of West Coast Steelhead. Federal Register 71FR834.

NMFS (National Marine Fisheries Service). 2010. Endangered and threatened wildlife and plants: threatened status for Southern Distinct Population Segment of eulachon. Federal Register 75FR13012.

NMFS (National Marine Fisheries Service). 2011. Anadromous Salmonid Passage Facility Design. NMFS, Northwest Region, Portland, Oregon.

NMFS (National Marine Fisheries Service). 2014. Endangered and threatened wildlife; final rule to revise the Code of Federal Regulations for species under the jurisdiction of the National Marine Fisheries Service. Federal Register 79FR20802.

NMFS SHIEER 2004, 70 FR 37160. June 28, 2005 - Final ESA listing determinations for 16 ESUs of West Coast salmon, and final 4(d) protective regulations for threatened salmonid ESUs; NMFS 2004. Salmonid Hatchery Inventory and Effects Evaluation Report (SHIEER). An evaluation of the effects of artificial propagation on the status and likelihood of extinction of west coast salmon and steelhead under the Federal Endangered Species Act. May 28, 2004. Technical Memorandum NMFS-NWR/SWR. U.S. Department of Commerce, National Oceanic and Atmospheric Administration, National Marine Fisheries Service. Portland, Oregon. 557p.

NPPC (Northwest Power Planning Council). 2001. Performance standards and indicators for the use of artificial production for anadromous and resident fish populations in the Pacific Northwest. Portland, Oregon. 19 pp.

Phinney, D. 2006. Compendium of Water Rights documents for Hatcheries and Wildlife areas. Washington Department of Fish and Wildlife Habitat Program. Olympia, Washington.

Piper, R., I.B. McElwain, L.E. Orme, J.P. McCraren, L.G. Fowler, J.R. Leonard, A.J. Trandahl, and V. Adriance. 1982. Fish Hatchery Management. United States Dept of Interior, Fish and Wildlife Service. Washington, D.C.

Seidel, P. 1983. Spawning guidelines for Washington Department of Fish and Wildlife hatcheries. Washington Department of Fish and Wildlife. Olympia, Washington.

Sharpe, C., P. Topping, T. Pearsons, J. Dixon and H. Fuss. 2008. Predation of naturally-produced fall Chinook fry by hatchery steelhead juveniles in Western Washington Rivers. Fish Program, Science Division Washington Department of Fish and Wildlife. Olympia, Washington.

Snow, C.G., A.R. Murdoch and T.H. Kahler. 2013. Ecological and demographic costs of releasing nonmigratory juvenile hatchery steelhead in the Methow River, Washington. North American Journal of Fisheries Management 33:6 1100-1112.

Steward, C. and T.C. Bjornn. 1990. Supplementation of salmon and steelhead stocks with hatchery fish; a synthesis of published literature. Idaho Cooperative Fish and Wildlife Research Unit. University of Idaho. Tech. Rpt. 90-1. Moscow, Idaho.

WDFW (Washington Department of Fish and Wildlife) and WWTIT (Western Washington Treaty Indian Tribes). 1998 (Updated 2006). Salmonid disease control policy of the fisheries Co-Managers of Washington State. Washington Department of Fish and Wildlife and Western Washington Treaty Indian Tribes, Olympia Washington.

WDFW (Washington Department of Fish and Wildlife). 2010. Draft Conservation and Sustainable Fisheries Plan (C&SFP). Washington Department of Fish and Wildlife. Olympia, Washington. 208 pp.

WDFW (Washington Department of Fish and Wildlife). 2010. WDFW Fisheries Management and Evaluation Plan (FMEP). Lower Columbia River. Submitted to NMFS Portland, Oregon.

WDFW (Washington Department of Fish and Wildlife). 2013. 2013 Future brood document. Washington Department of Fish and Wildlife. Olympia, Washington. Available from: <a href="http://wdfw.wa.gov/publications/01356/">http://wdfw.wa.gov/publications/01356/</a>.

WDFW (Washington Department of Fish and Wildlife). 2013. Hatcheries headquarters database. Hatcheries Data Unit, Washington Department of Fish and Wildlife. Olympia, Washington.

WDFW (Washington Department of Fish and Wildlife). 2013. Salmonid stock inventory (SaSI). Fish Program, Science Division. Washington Department of Fish and Wildlife. Olympia, Washington. Available from: http://wdfw.wa.gov/conservation/fisheries/sasi/.

WDFW (Washington Department of Fish and Wildlife). 2013. 2013/2014 Washington sport fishing rules. Washington Department of Fish and Wildlife. Olympia, Washington. Available from: http://wdfw.wa.gov/publications/01384/wdfw01384.pdf.

WDOE (Washington Department of Ecology). 2014. Water Resources Explorer. Retrieved July 8, 2014, from: <a href="https://fortress.wa.gov/ecy/waterresources/map/WaterResourcesExplorer.aspx">https://fortress.wa.gov/ecy/waterresources/map/WaterResourcesExplorer.aspx</a>.

Wipfli, M.S., J. Hudson, and J. Caouette. 1998 Influence of salmon carcasses on stream productivity: response of biofilm and benthic macroinvertebrates in southeastern Alaska, U.S.A. Can J. Fish. Aquat. Sci. 55: 1503-1511.

### Attachment 1: WDFW Virology Sampling 2006-2007 through 2012-2013: North Toutle Hatchery.

Hatchery/								Number of fish sampled						Cell		
Collection Site	Stock	Species	Date Sampled	Results	Comments	Life Stage	Sample number	OF	pools	K/S	pools	fry/visc	pools	ID	Line	Inoc Date
NO TOUTLE	TOUTLE R	FCHIN	10/10/06	PARAMYXOVIRUS	2+/12p K/S	AD	1011-1/2	60	12	60	12			F&P	С	
NO TOUTLE	TOUTLE R	SCOHO	11/14/06	NEV		AD	1115-31/32	60	12	60	12					
NO TOUTLE	TOUTLE R	FCHIN	10/15/07	PARAMYXOVIRUS	1+/12p K/S	AD	1016-8/9	60	12	60	12			F&P	С	
NO TOUTLE	TOUTLE R	SCOHO	11/06/07	NEV		AD	1107-11/12	60	12	60	12					
NO TOUTLE	TOUTLE R	FCHIN	05/12/08	NEV	RW2, RW3;diag; 10 <sup>0</sup> -10 <sup>-3</sup>	FRY/07	0513-3					10	2			
NO TOUTLE	TOUTLE R	FCHIN	10/20/08	NEV		AD	1021-13/14	60	12	60	12					
NO TOUTLE	TOUTLE R	SMCOHO	10/20/08	NEV		AD	1021-15/16	10	2	20	4					
NO TOUTLE	TOUTLE R	SCOHO	10/23/08	NEV		AD	1024-3/4	30	6	20	4					
NO TOUTLE	TOUTLE R	соно	10/29/08	NEV		AD	1029-12/13	10	2	20	4					
NO TOUTLE	TOUTLE R	СОНО	11/05/08	NEV		AD	1106-1	10	2							
NO TOUTLE	TOUTLE R	FCHIN	10/07/09	NEV		AD	1007-5/6	60	12	60	12					
NO TOUTLE	TOUTLE R	FCHIN	10/07/09	NEV		AD	1007-5/6	60	12	60	12					
NO TOUTLE	TOUTLE R	SCOHO	10/26/09	NEV		AD	1027-3/4	60	12	60	12					
NO TOUTLE	TOUTLE R	SCOHO	10/26/09	NEV		AD	1027-3/4	60	12	60	12					
NO TOUTLE	TOUTLE R	FCHIN	10/13/10	NEV		AD	1014-7/8	60	12	60	12					
NO TOUTLE	TOUTLE R	SCOHO	10/28/10	NEV		AD	1028-3/4	60	12	60	12					
NO TOUTLE	TOUTLE R	FCHIN	10/04/11	NEV		AD	1004-4/5	60	12	60	12					
NO TOUTLE	TOUTLE R	SCOHO	10/25/11	NEV		AD	1026-3/4	60	12	60	12					
NO TOUTLE	TOUTLE R	FCHIN	10/09/12	NEV		AD	1011-6/7	60	12	60	12		·			12/6/12
NO TOUTLE	TOUTLE R	NCOHO	10/24/12	NEV		AD	1024-6/7	60	12	60	12					4/24/13
NO TOUTLE	TOUTLE R	FCHIN	10/08/13	NEV		AD	1009-12/13	60	12	60	12		·			10/10/13
NO TOUTLE	TOUTLE R	scoho	10/23/13	NEV		AD	1024-1/2	60	12	60	12					10/24/13

Source: WDFW Fish Health Lab data (John Kerwin 2014)

### <u>Attachment 2: Fish health summaries: North Toutle Hatchery, July 1, 2008</u> through September 30, 2008 to October 1, 2001 through March 31, 2012...

### **North Toutle Hatchery S-Coho**

#### **Juveniles:**

### 2006 brood S-coho

Fish were healthy on release in May.

### 2007 brood S-coho

Green eggs were shipped to Grays River Hatchery for SAFE Deep River net pen program in November 2007. Fish were ponded in mid-March 2008; ponding was delayed due to cold temperatures. At the end of July 2008, all raceways were treated for *Columnaris* with TM-medicated feed. Loss was not significantly elevated. There were no problems during the rest of the reporting period. Fish were healthy on release in May.

### 2008 brood year S-coho

Fish were ponded in mid-March; ponding was delayed due to cold temperatures. Some fish lost due to deteriorating raceways (small fissures).at initial ponding. New bird netting minimized predation. A *Columnaris* outbreak during extremely warm temperatures during July and August 2009 caused high mortalities. Fry in April 2009 showed some anomalies in development due to temperature variation. Normal drop-out mortality occurred and no problems were seen afterwards through spring 2010. Fish were healthy on release in May 2010.

### 2009 brood year S-coho

No abnormal loss occurred during egg incubation. Green eggs were shipped to Grays River for the SAFE program in late-October. Cold temperatures during incubation contributed to a late ponding for these fish. Fish were ponded in mid-March. *Ichthyobodo* was found on gills early-June 2010, but didn't require treatment. *Columnaris* occurred in July 2010, and was successfully treated with TM medicated feed.

They exhibited good growth and very little mortality, except for an estimated 7,000 lost to otter predation. Fish were released healthy in May.

#### 2010 brood year S-coho

Green eggs were shipped to Grays River for the SAFE program. Fish were ponded in early-March. These fish remained healthy and did not require treatment for *Columnaris* during the mild summer. There was a normal amount of loss due to drop out and otter predation. Final rearing and release in November 2012.

#### 2011 brood S-coho

Fish were ponded in mid-March 2012 with no fish health issues observed. A portion was shipped to Grays River for the SAFE program.

### **Adults:**

<u>2009 S-coho</u> – Pre-spawning mortality was low in this stock. No viruses were detected in a sample of 60.

### 14 <u>SECTION 14. CERTIFICATION LANGUAGE AND SIGNATURE</u> <u>OF RESPONSIBLE PARTY</u>

"I hereby certify that the information provided is complete, true and correct to the best of my knowledge and belief. I understand that the information provided in this HGMP is submitted for the purpose of receiving limits from take prohibitions specified under the Endangered Species Act of 1973 (16 U.S.C.1531-1543) and regulations promulgated thereafter for the proposed hatchery program, and that any false statement may subject me to the criminal penalties of 18 U.S.C. 1001, or penalties provided under the Endangered Species Act of 1973."

Name, Title, and Signature of Applicant:	
Certified by	_ Date:

### 15 ADDENDUM A. PROGRAM EFFECTS ON OTHER (AQUATIC OR TERRESTRIAL) ESA-LISTED POPULATIONS. (Anadromous

salmonid effects are addressed in Section 2).

## 15.1 List all ESA permits or authorizations for USFWS ESA-listed, proposed, and candidate salmonid and non-salmonid species associated with the hatchery program.

The WDFW and the USFWS have a Cooperative Agreement pursuant to section 6(c) of the Endangered Species Act that covers the majority of the WDFW actions, including hatchery operations.

"The department is authorized by the USFWS for certain activities that may result in the take of bull trout, including salmon/steelhead hatchery broodstocking, hatchery monitoring and evaluation activities and conservation activities such as adult traps, juvenile monitoring, spawning ground surveys..."

### 15.2 Describe USFWS ESA-listed, proposed, and candidate salmonid and non-salmonid species and habitat that may be affected by hatchery program.

Several listed and candidate species are found in Cowlitz, Clark and Skamania Counties; however the hatchery operations and facilities for this program do not fall within the critical habitat for any of these species. As such there are no effects anticipated for these species.

"No effect" for the following listed species:

Bull trout (Salvelinus confluentus) – Threatened (Critical Habitat Designated)

Northern Spotted owl (Strix occidentalis caurina) – Threatened (Critical Habitat Designated)

Marbled murrelet (*Brachyramphus marmoratus*) –Threatened (Critical Habitat Designated)

Columbian White-Tailed deer (Odocoileus virginianus leucurus) – Endangered

Canada Lynx (Lynx canadensis) –Threatened

Grizzly bear (Ursus arctos horribilis) - Threatened

Gray Wolf (Canis lupus) - Threatened

Nelson's checker-mallow (Sidalcea nelsoniana) - Threatened

Bradshaw's desert-parsley (Lomatium bradshawii) - Endangered

Water howellia (Howellia aquatilis) –Threatened

Golden Paintbrush (Castilleja levisecta) [historic]

#### **Candidate Species:**

Fisher (Martes pennanti) – West Coast DPS

Mardon skipper (*Polites mardon*)

Oregon spotted frog (Rana pretiosa)

Whitebark pine (*Pinus albicaulis*)

North American wolverine (Gulo gulo luteus) – contiguous U.S. DPS

(Brush Prairie) Mazama pocket gopher (Thomomys mazama ssp. oregonus)

### 15.3 Analyze effects.

Not applicable.

#### 15.4 Actions taken to minimize potential effects.

Program coho are released fully smolted to foster rapid outmigration from the basin and to minimize predation and residualism risks.

#### 15.5 References

Not applicable.

### 16 "Take" Tables

Table 1. Estimated listed salmonid take levels of by hatchery activity

Table 1. Estimated listed samfoling take levels of by flatenery activity.										
Listed species affected:	ESU/Po	ESU/Population:			Activity:					
Spring Chinook (Oncorhynchus tshawytscha)	Lower C	Lower Columbia River Spring Chinook			North Toutle Type-S Coho Program					
Location of hatchery activity:	Dates of	Cactivity:		Hatchery pr	ogram operator:					
North Hatchery, Green River (WRIA 26.0323) at RM 0.81	Septemb	er-November		WDFW						
Tributary to the NF Toutle River (WRIA 26.0314) at R.M. 11.5										
Type of Take		Annual Take of Listed Fish By Life Stage (Number of Fish)								
		Egg/Fry	Juvenile	/Smolt	Adult	Carcass				
Observe or harass <sup>a</sup>										
Collect for transport <sup>b</sup>										
Capture, handle, and release c					TBD					
Capture, handle, tag/mark/tissue sample, and released <sup>d</sup>										
Removal (e.g. broodstock) <sup>e</sup>				TBD						
Intentional lethal take <sup>f</sup>				TBD						
Unintentional lethal take <sup>g</sup>		TB	D							
Other Take (specify) h										

Take Table to be submitted to NOAA-NMFS, in progress.

- a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
- b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
- c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
- d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
- e. Listed fish removed from the wild and collected for use as broodstock.
- f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
- g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
- h. Other takes not identified above as a category.

#### Instructions:

- 1. An entry for a fish to be taken should be in the take category that describes the greatest impact.
- 2. Each take to be entered in the table should be in one take category only (there should not be more than one entry for the same sampling event).
- 3. If an individual fish is to be taken more than once on separate occasions, each take must be entered in the take table.

**Table 2.** Estimated listed salmonid take levels of by hatchery activity.

Listed species affected:		ESU/Population:			Activity:					
Fall Chinook (Oncorhynchus tshawytscha)	Lower C	ower Columbia River Fall Chinook			North Toutle Hatchery Type-S Coho Program					
Location of hatchery activity:		ates of activity:			Hatchery program operator:					
North Hatchery, Green River (WRIA 26.0323) at RM 0.81	Septemb	September-November			WDFW					
Tributary to the NF Toutle River (WRIA 26.0314) at R.M. 11.5										
Type of Take		Annual Take of Listed Fish By Life Stage (Number of Fish)								
		Egg/Fry	Juvenile/Smolt		Adult	Carcass				
Observe or harass <sup>a</sup>										
Collect for transport <sup>b</sup>	Collect for transport <sup>b</sup>									
Capture, handle, and release c				TBD						
Capture, handle, tag/mark/tissue sample, and released <sup>d</sup>										
Removal (e.g. broodstock) <sup>e</sup>										
Intentional lethal take <sup>f</sup>				TBD						
Unintentional lethal take <sup>g</sup>		TBI	)	TBD	_					
Other Take (specify) h										

- a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
- b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
- c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
- d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
- e. Listed fish removed from the wild and collected for use as broodstock.
- f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
- g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
- h. Other takes not identified above as a category.

#### Instructions:

- 1. An entry for a fish to be taken should be in the take category that describes the greatest impact.
- 2. Each take to be entered in the table should be in one take category only (there should not be more than one entry for the same sampling event).
- 3. If an individual fish is to be taken more than once on separate occasions, each take must be entered in the take table.

**Table 3.** Estimated listed salmonid take levels of by hatchery activity.

Listed species affected:	opulation: Activity:									
Steelhead (Oncorhynchus mykiss)	Lower C	olumbia River Steelhead		North Toutle Hatche	ry Type-S Coho Program					
Location of hatchery activity:		activity: Hatchery program operator:								
North Hatchery, Green River (WRIA 26.0323) at RM 0.81	Septemb	er-November		WDFW						
Tributary to the NF Toutle River (WRIA 26.0314) at R.M. 11.5										
Type of Take		Annual Take of Listed Fish By Life Stage (Number of Fish)								
		Egg/Fry	Juvenile/Smolt		Adult	Carcass				
Observe or harass <sup>a</sup>										
Collect for transport b										
Capture, handle, and release <sup>c</sup>					TBD					
Capture, handle, tag/mark/tissue sample, and released <sup>d</sup>										
Removal (e.g. broodstock) <sup>e</sup>										
Intentional lethal take <sup>f</sup>										
Unintentional lethal take <sup>g</sup>			TBD							
Other Take (specify) h										

<sup>\*</sup> Highest number of unmarked winter-late steelhead trapped and released in the Lewis River.

- a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
- b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
- c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
- d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
- e. Listed fish removed from the wild and collected for use as broodstock.
- f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
- g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
- h. Other takes not identified above as a category.

**Table 4.** Estimated listed salmonid take levels of by hatchery activity.

Listed species affected:	Listed species affected: ESU/Popu			Activity:	ctivity:					
Coho ( <i>Oncorhynchus kisutch</i> )				th Toutle Hatchery Type-S Coho Program						
Location of hatchery activity:				Hatchery program operator:						
	North Hatchery, Green River (WRIA 26.0323) at RM 0.81 September			WDFW						
Tributary to the NF Toutle River (WRIA 26.0314) at R.M. 11.5										
Type of Take		Annual Take of Listed Fish By Life Stage ( <u>Number of Fish</u> )								
		Egg/Fry	Juvenile/Smolt		Adult	Carcass				
Observe or harass <sup>a</sup>										
Collect for transport b										
Capture, handle, and release <sup>c</sup>				TBD						
Capture, handle, tag/mark/tissue sample, and released <sup>d</sup>										
Removal (e.g. broodstock) <sup>e</sup>				TBD						
Intentional lethal take <sup>f</sup>	TBD	TBD		TBD						
Unintentional lethal take <sup>g</sup>		TBD								
Other Take (specify) h										

<sup>\*</sup> Based on 90% egg-to-fry survival, and 90% fry-to-smolt survival.

- a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
- b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
- c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
- d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
- e. Listed fish removed from the wild and collected for use as broodstock.
- f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
- g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
- h. Other takes not identified above as a category.

**Table 5.** Estimated listed salmonid take levels of by hatchery activity.

Listed species affected:	opulation: Activity			vity:					
Chum (Oncorhynchus keta)	Columbi	umbia River Chum Nor			North Toutle Hatchery Type-S Coho Program				
Location of hatchery activity:		of activity: Hatch			rogram operator:				
North Hatchery, Green River (WRIA 26.0323) at RM 0.81	Septemb	er-November		WDFW					
Tributary to the NF Toutle River (WRIA 26.0314) at R.M. 11.5									
Type of Take		Annual Take of Listed Fish By Life Stage (Number of Fish)							
		Egg/Fry	Juvenile/S	molt	Adult	Carcass			
Observe or harass <sup>a</sup>									
Collect for transport <sup>b</sup>									
Capture, handle, and release c									
Capture, handle, tag/mark/tissue sample, and released <sup>d</sup>									
Removal (e.g. broodstock) <sup>e</sup>									
Intentional lethal take <sup>f</sup>									
Unintentional lethal take <sup>g</sup>		TBD							
Other Take (specify) h									

- a. Contact with listed fish through stream surveys, carcass and mark recovery projects, or migrational delay at weirs.
- b. Take associated with weir or trapping operations where listed fish are captured and transported for release.
- c. Take associated with weir or trapping operations where listed fish are captured, handled and released upstream or downstream.
- d. Take occurring due to tagging and/or bio-sampling of fish collected through trapping operations prior to upstream or downstream release, or through carcass recovery programs.
- e. Listed fish removed from the wild and collected for use as broodstock.
- f. Intentional mortality of listed fish, usually as a result of spawning as broodstock.
- g. Unintentional mortality of listed fish, including loss of fish during transport or holding prior to spawning or prior to release into the wild, or, for integrated programs, mortalities during incubation and rearing.
- h. Other takes not identified above as a category.